We studied 2332 individuals with monoallelic mutations in MUTYH among 9504 relatives of 264 colorectal cancer (CRC) cases with a MUTYH mutation. We estimated CRC risks through 70 years of age of 7.2% for male carriers of monoallelic mutations (95% confidence interval [CI], 4.6%–11.3%) and 5.6% for female carriers of monoallelic mutations (95% CI, 3.6%–8.8%), irrespective of family history. For monoallelic MUTYH mutation carriers with a first-degree relative with CRC diagnosed by 50 years of age who does not have the MUTYH mutation, risks of CRC were 12.5% for men (95% CI, 8.6%–17.7%) and 10% for women (95% CI, 6.7%–14.4%). Risks of CRC for carriers of monoallelic mutations in MUTYH with a first-degree relative with CRC are sufficiently high to warrant more intensive screening than for the general population.

Keywords: Colon Cancer; Genetics; Base Excision Repair Gene; DNA Damage Response.

**Results**

We identified 9504 relatives (4613 females) from the families of the 264 (236 population-based and 28 clinic-based) probands with a monoallelic or biallelic MUTYH mutation from the Colon Cancer Family Registry; 138 (52%) from the United States, 81 (31%) from Canada, and 45 (17%) from Australia and New Zealand. In the relatives, we observed 261 with CRC (114 were female with median age increased risk of colorectal cancer (CRC)). Individuals with germline mutations in one allele (monoallelic mutation carriers) have a small increased risk of CRC. Due to the rarity of these mutations, previous studies have had limited ability to provide precise estimates of age- and sex-specific CRC risks for MUTYH mutation carriers. In addition, the variability in CRC risk between carriers has not been quantified. Modeling of this variability can indicate a potential role for modifiers of risk.

**Abbreviations used in this paper:** CI, confidence interval; CRC, colorectal cancer; HR, hazard ratio.

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MUTYH mutation status was known for 340 relatives (13 biallelic mutation carriers, 142 monoallelic mutation carriers, and 185 noncarriers). We estimated an additional 43 biallelic and 2190 monoallelic mutation carriers among non-genotyped relatives, giving a total estimated number of 56 biallelic and 2332 monoallelic mutation-carrying relatives in our sample.

Our methods allowed for CRC risk estimation in mutation families to be due to the MUTYH mutation, as well as polygenic factors (combination of a large number of CRC-associated genetic susceptibility loci).7 We estimated CRC risks through 70 years of age for male and female to be: 75.4% (95% confidence interval [CI]: 41.2%–96.6%) and 71.7% (95% CI, 44.5%–92.1%), respectively, for biallelic mutation carriers, and 7.2% (95% CI, 4.6%–11.3%) and 5.6% (95% CI, 3.6%–8.8%), respectively, for monoallelic mutation carriers (Figure 1). The estimated CRC risks through 70 years of age for monoallelic mutation carriers with a first-degree relative with CRC were similar whether the relative was untested or a noncarrier or a monoallelic mutation carrier: approximately 12% (95% CI, 9%–18%) and 10% (95% CI, 7%–14%) for males and females, respectively, in comparison with males and females from the general population (2.9% and 2.1%, respectively). However, if their affected first-degree relative was a biallelic mutation carrier, then risks of CRC through 70 years of age, for monoallelic mutation carriers was estimated to be 10.4% (95% CI, 7.0%–15.0%) and 8.2% (95% CI, 5.4%–12.0%) for males and females, respectively (Table 1). In addition, we estimated CRC risks for 6 other scenarios (Supplementary Figure 1). The highest risk of CRC for a monoallelic mutation carrier corresponded to having 2 affected first-degree relatives; 1 is a biallelic mutation carrier and 1 is a noncarrier (Supplementary Figure 1C).

We found no evidence for a difference in hazard ratios (HRs) of CRC for biallelic mutation carriers between males and females (HR, 108; 95% CI, 25.9–454 vs HR, 129; 95% CI, 43.7–380; P = .85), or for monoallelic mutation carriers between males and females (HR, 2.46; 95% CI, 1.54–3.93 vs HR, 2.67; 95% CI, 1.67–4.26; P = .81). Hazard ratio of CRC for Y179C monoallelic carriers was higher than for G396D monoallelic carriers (HR, 4.81; 95% CI, 3.00–7.71 vs HR, 2.42; 95% CI, 1.48–3.98; P = .05), but there was no difference between biallelic carriers of Y179C and G396D (P = .84) (Supplementary Table 1).

The standard deviation of the polygenic component was estimated to be 1.11 (95% CI, 0.74–1.49; P < .001); see the Supplementary Material for a general formula relating this standard deviation to the HR. At ages younger than 50 years, this formula reduces to Pharoah’s formula for early-onset disease7 and says that monoallelic MUTYH mutation carriers with an affected first-degree relative have CRC incidences approximately 4.58 (for males) or 4.97 (for females) times the population incidences. However, Supplementary Figure 2 gives precise HRs for all ages and shows that by age 70 years, Pharoah’s formula overestimates relative risks by roughly 30%.

**Discussion**

Our finding of almost complete penetrance for biallelic MUTYH mutation carriers is consistent with previous studies.7–10 There is some evidence that biallelic mutation carriers move rapidly along a mutator phenotype progression to cancer.11 These findings support the recommendation that biallelic mutation carriers should consider prophylactic total colectomy with ileorectal anastomosis, depending on the individual, age of presentation, and number and size of polyps present.12
We estimated monoallelic mutation carriers had, on average, an approximately 2.5-fold increased risk of CRC compared with the general population, consistent with one previous study. This level of increased risk for monoallelic mutation carriers is similar to that for people with a first-degree relative with CRC, who are recommended colonoscopy every 5 years starting 10 years younger than the youngest case in the family and before age 50 years. However, monoallelic mutation carriers who have an affected first-degree relative were at approximately 5-fold increased risk. For these carriers, colonoscopy beginning at age 40 years, with follow-up at intervals dependent on the presence or absence of polyps but no less often than every 5 years, may be reasonable.

We observed strong evidence that CRC risks for carriers are highly heterogeneous. The observed heterogeneity in risk could also be caused by environmental factors shared between family members or by differences in risk between mutations. To our knowledge, so far the only study investigating modifiers of CRC risks for MUTYH mutation carriers was on the relationship with hormone-replacement therapy, which reported no evidence of interaction between hormone-replacement therapy and MUTYH mutations.

In this study of 12 variants of MUTYH mutations, 93% of the MUTYH mutations were Y179C and G396D (Supplementary Table 2); consistent with a previous study of Caucasians. We found CRC risk was higher for monoallelic carriers of Y179C than for G396D, which is consistent with previous studies. However, given our approach of genotyping for 12 mutations by mass spectrometry and WAVE (Transgenomic, Omaha, NE), followed by confirmatory Sanger sequencing of MUTYH in carriers (see Materials and Methods in Supplementary Material), there is the possibility that we missed other pathogenic mutations in MUTYH that were not 1 of the 12 mutations genotyped. Although we identified additional variants from Sanger sequencing, their pathogenicity was considered inconclusive (unclassified variants) and therefore not included in this analysis. Additional MUTYH mutations might reside in different ethnic groups, however, this cohort was predominantly Caucasian.

We used sophisticated statistical techniques to adjust for ascertainment, to account for residual familial aggregation of disease and therefore avoid bias, and to use data for all family members, whether genotyped or not, and therefore maximized statistical power and avoided survival bias.

In conclusion, using the largest international study, to date we have produced unbiased estimates of CRC risks for MUTYH mutation carriers, which are the most precise and reliable currently available. In addition to the confirmed very high risk of CRC to biallelic MUTYH mutation carriers, CRC risk for monoallelic mutation carriers depends on family history and can be sufficiently high to warrant consideration of more intensive CRC screening than for the general population.

### Supplementary Material

Note: To access the supplementary material accompanying this article, visit the online version of Gastroenterology at [www.gastrojournal.org](http://www.gastrojournal.org), and at [http://dx.doi.org/10.1053/j.gastro.2014.01.022](http://dx.doi.org/10.1053/j.gastro.2014.01.022).

### References


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Conflicts of interest
The authors disclose no conflicts.

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The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of the collaborating centers in the Cancer Family Registries, nor does mention of trade names, commercial products, or organizations imply endorsement by the US Government or the Cancer Family Registry. Authors had full responsibility for the design of the study, the collection of the data, the analysis and interpretation of the data, the decision to submit the manuscript for publication, and the writing of the manuscript.
Supplementary Material and Methods

Study Sample

Subjects were from families for which at least 1 family member had been identified as carrying a germline monoallelic or biallelic mutation in MUTYH. These families were recruited by the Colon Cancer Family Registry via probands between 1997 and 2007. Population-based probands were recently diagnosed colorectal cancer cases from state or regional population cancer registries in the United States (Washington, California, Arizona, Minnesota, Colorado, New Hampshire, North Carolina, and Hawaii), Australia (Victoria) and Canada (Ontario). Clinic-based probands were enrolled from multiple-case families referred to family cancer clinics in the United States (Mayo Clinic, Rochester, Minnesota, and Cleveland Clinic, Cleveland, Ohio), Australia (Melbourne, Adelaide, Perth, Brisbane, Sydney) and New Zealand (Auckland). No cases were ascertained because of having polyps alone. All cases with familial adenomatous polyposis were excluded.

Probands were asked for permission to contact their relatives to seek their enrollment in the Cancer Family Registry. For population-based families, first-degree relatives of probands were recruited at all centers and recruitment was extended to more distant relatives at some centres. For clinic-based families, recruitment was based on availability but attempts were made to recruit up to second-degree relatives of affected individuals (detail in Newcomb et al\(^1\)). Written informed consent was obtained from all study participants, and the study protocol was approved by the Institutional Research Ethics Review Board at each center.

Data Collection

Standardized questionnaires were used to collect information on demographics, personal characteristics, personal and family history of cancer, cancer screening history, history of polyps, polypectomy and other procedures were obtained by questionnaires from all probands and participating relatives. Reported cancer diagnoses and age at diagnosis were confirmed, where possible using pathology reports, medical records, cancer registry reports, and/or death certificates. Standardized protocols were used to collect and prepare blood samples and tumour tissues for genetic testing and laboratory analyses.

MUTYH Mutation Testing

We tested all probands for mutations in the MUTYH gene, and also relatives of those with mutations who provided a DNA sample. As described in detail by Cleary et al\(^2\), genomic DNA extracted from each participant was sent to a central testing facility (Analytic Genetics Technology Centre, Toronto, Canada). DNA was screened for 12 previously identified MUTYH mutations: Y179C, G396D, Y104X, R274Q, E480X, Q391X, c.1145delG, c.933+3A>C, c.1437_1439delGGA, R241W, c.1228_1229insGG, and c.1187-2A→G using the MassArray MALDI-TOF Mass Spectrometry system (Sequenom, San Diego, CA). Screening for R241W, c.1228_1229insGG, and c.1187-2A→G was discontinued when testing of 6000 samples failed to identify any carriers of these 3 variants. All samples with mass spectrometry mobility shifts underwent screening of the entire MUTYH coding region, promoter, and splice sites regions by denaturing high-performance liquid chromatography (Transgenomic Wave 3500HT System; Transgenomic) to confirm the mutation and to identify additional mutations. All mass spectrometry–detected variants and WAVE mobility shifts were submitted for sequencing for mutation confirmation (ABI PRISM 3130XL Genetic Analyzer; Applied Biosystems, Carlsbad, CA).

From the Colon Cancer Family Registry, we identified 273 probands who were known to carry germline mutations in MUTYH. We excluded 9 probands who were also known to carry pathogenic mismatch repair germline mutations in a mismatch repair gene (Lynch syndrome). Of the remaining 264 probands, 41 were biallelic mutation carriers and 223 were monoallelic mutation carriers. The variants of MUTYH mutations of the probands are shown in Supplementary Table 2. The mean age at diagnosis of colorectal cancer was 47.7 years (SD 10.0 years) in probands with biallelic mutations and 52.0 years (SD 11.9 years) in probands with monoallelic mutations.

Statistical Analysis

The median, range, mean, and SD of the age at colorectal cancer diagnosis were calculated using Stata 12.1 software (StataCorp, College Station, TX, 2011). HRs, that is, the age- and sex-specific cancer incidence for carriers divided by that for the general population,\(^3\) were estimated using modified segregation analysis.\(^4,5\) Models were fit by maximum likelihood with the statistical package MENDEL version 3.2.\(^6\) Estimates were appropriately adjusted for the clinic- and population-based ascertainment of families using a combination of retrospective likelihood and ascertainment-corrected joint likelihood.\(^4,7–9\) in which each pedigree’s data were conditioned on the proband’s genotype, cancer status, and age of onset (for population-based families) or on the proband’s genotype and the affected statuses and ages of onset of all family members at the time the proband was found to be a MUTYH mutation carrier (for clinic-based families).

To model any residual familial aggregation of colorectal cancer risk, a mixed model that incorporated an unmeasured polygene in addition to the major gene,\(^6,10\) was used in the modified segregation analyses (see detail methods in a previous report\(^11\)). This mixed model was used because major gene models (which attribute all familial aggregation to the major gene being studied alone) are often biased.\(^8\) The polygenic part of this mixed model, which models the cumulative effect of a large number of biallelic genes that individually have small effects on cancer susceptibility, was implemented as a hypergeometric polygenic model with 4 loci.\(^4,11\) Under this model, the number of disease alleles for each person is approximately normally distributed and is correlated within families with correlation coefficients equal to the kinship coefficients.\(^9\)

Estimated cumulative risks (penetrance) of cancers to age \(\tau\) years for carriers living in the United States were calculated
by from the relevant population incidences $\lambda_0(\tau)$ at age $\tau$ years multiplied and the estimated HR $\theta$ with the formula:

$$1 - \exp\left(-\int_0^\tau \theta \lambda_0(\tau) d\tau\right)$$

We estimated the total number of carriers in the study using the same method in previous studies\textsuperscript{12,13}; by summing \textit{MUTYH} carrier probabilities for all individuals, as calculated from Mendel’s laws of inheritance, the known genetic relationship of each individual to his or her genotyped relatives (but not affected status) and a population allele frequency of 0.0085. These calculations were performed using R 2.15.0 (R Foundation for Statistical Computing, Vienna, Austria, 2008) and a modified version of Mendel 3.2.\textsuperscript{6}

Observation time for each subject started at birth and ended at first diagnosis of colorectal cancer or other cancer, first polypectomy, last follow-up, or death, whichever occurred first. Where age at diagnosis of cancer was not reported ($n = 43$; 16% of all cancer cases), we assumed the age of diagnosis to be 1 year before the last known age or, if last known age was not available, the median age at diagnosis of CRC for the general population obtained from Surveillance, Epidemiology, and End Results Cancer Statistics Review (1975–2007).\textsuperscript{14}

**Effect of Family History on Disease Risks for Major Gene Mutation Carriers**

In this section, we describe statistical methods used to derive the age-dependent cumulative risks and HRs for a carrier of a major gene mutation who has an affected first-degree relative, based on the genetic mixed model used in this article. The cumulative risks and HRs are given in Equations (2) and (3), respectively, and Equation (4) gives approximate HRs for rare diseases or for early-onset forms of common diseases.

Consider 2 individuals who are indexed by $i = 1$ and 2 and let $T_i, G_i$ and $H_i$ be random variables representing (respectively) the age at disease onset, the major gene genotype and the polygenotype of individual $i$. We assume the 2 individuals are first-degree relatives, so that $(H_1, H_2)$ is a bivariate, normally distributed random variable, with the correlation coefficient of $H_1$ and $H_2$ being $\frac{1}{2}$ and each $H_i$ having mean $\mu$ and variance $\sigma^2$. For simplicity, we assume that mutations in the major gene are rare, so that we can ignore biallelic carriers and we can assume $P(G_2 = 1|G_1 = 1) = \frac{1}{2}$ where $G_i = 0$ if $i$ is a noncarrier and $G_i = 1$ if $i$ is a carrier, although the derivation can easily be changed to incorporate common alleles (and general modes of inheritance). We also assume $T_1$ and $T_2$ are conditionally independent given all genotypes and polygenotypes, that $T_1$ only depends on $G_1$ and $H_1$ (and similarly for $T_2$) and that $G_1$ and $G_2$ are independent of $H_1$ and $H_2$.

Let $\lambda_0(t)$ be the average incidence of disease at age $t$ years for noncarriers (the average being over the polygene) and recall that the hazard function $\lambda(x)$ of any continuous random variable $X$ is defined to be:

$$\lambda(x) = -\frac{d}{dx}\log(1 - P(X \leq x)).$$

Then, as in the rest of this article, we assume that the hazard of $T_i$ (ie, the incidence of disease for individual $i$) at age $t_i$ years, conditioned on $G_i = g_i$ and $H_i = h_i$, is equal to $\theta_{gh_i} \lambda_0(t_i)$, where $\theta_{gh_i}$ is the HR associated with major gene genotype $g_i$ and $r_i = e^{\delta_i}$ is the HR associated with polygenotype $h_i$. Using the shorthand $t_i, g_i$ and $h_i$ for the events $T_i \leq t_i, G_i = g_i$ and $H_i = h_i$ (respectively) in all probability statements from now on, it therefore follows from (1) that:

$$P(t_i|g_i, h_i) = 1 - s_i^{[t_i]}$$

where $s_i = \exp(-\int_0^t \lambda_0(\tau)d\tau)$ is the survival function to age $t_i$ years for noncarriers. We assume the HR $\theta_0$ for noncarriers is 1, so in order that the average incidence of disease for noncarriers at age $t$ years will equal $\lambda_0(t)$, we need each log-normal random variable $e^{\delta_i}$ to have an expected value of 1, ie, we need $\mu = -\sigma^2/2$.

We are interested in the cumulative risk of disease for individual 1 by age $t_1$ given that he or she is a carrier and that his or her first-degree relative, individual 2, was affected by age $t_2$. In other words, we want to calculate $P(t_1|g_1, t_2)$ when $g_1 = 1$. But $P(t_1|g_1, t_2) = P(t_1|g_1, t_2)/P(t_2|g_1)$ and

$$P(t_1, t_2|g_1) = \sum_{g_2, h_2} P(t_1, t_2, g_2, h_1, h_2|g_1) = \sum_{g_2, h_2} P(t_1, t_2|g_1, g_2, h_1, h_2)P(g_2, h_1, h_2|g_1)$$

$$= \sum_{g_2, h_2} P(t_1|g_1, h_1)P(t_2|g_2, h_2)P(g_2|g_1)P(h_1, h_2)$$

$$= \frac{1}{2^2} \sum_{h_1, h_2} P(h_1, h_2)((1 - s_1^{[t_1]})(1 - s_2^{[t_2]}) + (1 - s_1^{[t_1]})(1 - s_2^{[t_2]}))$$

$$= \frac{1}{2} \mathbb{E}[(1 - s_1^{[t_1]})(2 - s_2^{[t_2]} - s_2^{[t_2]}),$$
where \( R_1 = e^{R_1} \), \( \mathbb{E} \) is the expectation functional, and \( \theta = \theta_1 \) is the HR for carriers of major gene mutations compared with noncarriers. Similarly, \( P(t_2|g_1) = \frac{1}{2} \mathbb{E}[2 - s_r^2 - s_s^2] \), so the cumulative risk is:

\[
P(t_1|g_1, t_2) = \frac{\mathbb{E}[(1 - s_1^{\theta R_1})(2 - s_r^2 - s_s^2)]}{\mathbb{E}[2 - s_r^2 - s_s^2]} \\
= 1 - \frac{\mathbb{E}[s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]}{\mathbb{E}[2 - s_r^2 - s_s^2]} \tag{2}
\]

By (1), the hazard \( \lambda(t_1|g_1, t_2) \) of \( T_1 \) at \( t_1 \) given \( G_1 = 1 \) and \( T_2 \leq t_2 \) is therefore:

\[
\lambda(t_1|g_1, t_2) = \frac{d}{dt_1} \log(1 - P(t_1|g_1, t_2)) \\
= \frac{d}{dt_1} \log \mathbb{E}[s_1^{\theta R_1}(2 - s_r^2 - s_s^2)] \\
= -\frac{\mathbb{E}[d(\theta R_1 \log s_1)(2 - s_r^2 - s_s^2)]}{\mathbb{E}[s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]} \\
= \frac{-\mathbb{E}[d(\log s_1)\theta R_1 s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]}{\mathbb{E}[s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]} \\
= \theta \lambda_0(t_1) \frac{\mathbb{E}[R_1 s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]}{\mathbb{E}[s_1^{\theta R_1}(2 - s_r^2 - s_s^2)]}.
\]

So the HR \( \text{HR}(t_1|g_1, t_2) \) for \( T_1 \) given \( G_1 = 1 \) and \( T_2 \leq t_2 \) (i.e., the relative risk of disease at age \( t_1 \) for carriers with an affected first-degree relative) is:

\[
\text{HR}(t_1|g_1, t_2) = \theta e^{\theta R_1 s_1^{\theta R_1}(2 - s_r^2 - s_s^2)}.
\]

It is unlikely that a simple formula can be given for these expectations because, for instance, \( \mathbb{E}[s_1^{\theta R_1}] = M_{\theta R_1}(\theta \log s_1) \) where \( M_{\theta R_1} \) is the moment-generating function of the log-normal random variable \( R_1 \), and no closed-form expression for \( M_{\theta R_1} \) is known. However, the expectations can be readily evaluated by simulating draws of \( (H_1, H_2) \) from the multivariate normal distribution described here.

We can also derive approximations to these expectations if we restrict our attention to rare diseases or to early-onset forms of common diseases. For in these cases, each \( s_i \approx 1 \) so \( |\log s_i| < < 1 \) and we can evaluate approximations to the above expectations to a first order in \( \log s_i \). Using \( s_i^{\theta R_i} = e^{\theta R_i \log s_i} \approx 1 + \theta R_i \log s_i \) we have

\[
\mathbb{E}[\theta R_i s_i^{\theta R_i}(2 - s_r^2 - s_s^2)] \\
\approx [2(1 + \theta R_i \log s_i) - (1 + \theta R_1 \log s_1 + \theta R_2 \log s_2)] \\
= -(\theta + 1)\log s_2 \mathbb{E}[R_2] = -(\theta + 1)\log s_2.
\]

Similarly, \( \mathbb{E}[R_1 s_1^{\theta R_1}(2 - s_r^2 - s_s^2)] \approx -(\theta + 1)\log s_2 \mathbb{E}[R_1 R_2] \).

But \( R_1 R_2 = e^{R_1 + R_2} \) is log-normally distributed and \( H_1 + H_2 \) has mean \( 2\mu = -\sigma^2 \) and variance \( 3\sigma^2 \), so \( \mathbb{E}[R_1 R_2] = e^{\sigma^2/2} \).

Therefore

\[
\text{HR}(t_1|g_1, t_2) \approx \theta e^{\sigma^2/2} \tag{4}
\]

to first order in \( \log s_1 \) and \( \log s_2 \). Setting \( \theta \), the major gene HR, equal to 1 then gives the formula derived in the appendix of Pharoah et al.\(^{15}\)

---

**Supplementary References**

### Supplementary Table 1. HR (95% CI) of CRC for Biallelic and Monoallelic MUTYH Mutation Carriers

<table>
<thead>
<tr>
<th>Sex</th>
<th>Biallelic mutation carriers</th>
<th>Monoallelic mutation carriers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male</td>
<td>108 (25.9–454)</td>
<td>2.46 (1.54–3.93)</td>
</tr>
<tr>
<td>Female</td>
<td>129 (43.7–380)</td>
<td>2.67 (1.67–4.26)</td>
</tr>
<tr>
<td>Variants</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Y179C</td>
<td>115 (23.3–569)</td>
<td>4.81 (3.00–7.71)</td>
</tr>
<tr>
<td>G396D</td>
<td>94.2 (29.2–304)</td>
<td>2.42 (1.48–3.98)</td>
</tr>
</tbody>
</table>

**NOTE.** Values are HR (95% CI).

### Supplementary Table 2. Variants of MUTYH Mutations in Probands

<table>
<thead>
<tr>
<th>Variant 1</th>
<th>Variant 2</th>
<th>No. of probands</th>
</tr>
</thead>
<tbody>
<tr>
<td>Biallelic carriers</td>
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</tr>
<tr>
<td>1187G&gt;A, GGT&gt;GAT, G396D</td>
<td>1187G&gt;A, GGT&gt;GAT, G396D</td>
<td>14</td>
</tr>
<tr>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>6</td>
</tr>
<tr>
<td>1187G&gt;A, GGT&gt;GAT, G396D</td>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>10</td>
</tr>
<tr>
<td>1187G&gt;A, GGT&gt;GAT, G396D</td>
<td>821G&gt;A, CGG&gt;CAG, R274Q</td>
<td>1</td>
</tr>
<tr>
<td>1187G&gt;A, GGT&gt;GAT, G396D</td>
<td>933+3A&gt;C, splicesite</td>
<td>2</td>
</tr>
<tr>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>933+3A&gt;C, splicesite</td>
<td>1</td>
</tr>
<tr>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>734G&gt;A, CGT&gt;CAG, R245H</td>
<td>2</td>
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<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>312C&gt;A, TAC&gt;TAA, Y104X</td>
<td>1</td>
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<tr>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>1437_1439delGGA, in-frame deletion</td>
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<tr>
<td>536A&gt;G, TAC&gt;TGC, Y179C</td>
<td>1145delC</td>
<td>2</td>
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<tr>
<td>1171C&gt;T, CAG&gt;TAG, Q391X</td>
<td>1437_1439delGGA, in-frame deletion</td>
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<td>Monoallelic carriers</td>
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Supplementary Figure 1. Cumulative risk of CRC to age 70 years for counselees in 6 different scenerios (A–F). We assumed that CRC to be diagnosed at or before age 70 years for affected persons, and last known age to be age 70 years for unaffected persons. +/+, biallelic MUTYH mutation carrier; +/-, monoallelic MUTYH mutation carrier; --, no MUTYH mutation; ?, ungenotyped for MUTYH; filled in symbol, CRC; <70, age of CRC diagnosis (years); arrow, counselee.

Supplementary Figure 2. Age-specific HRs for monoallelic mutation carriers with affected first-degree relatives (FDRs) diagnosed by certain ages (gray lines), as well as the HRs given in Equation 4 of the Supplementary Materials and Methods and derived under an early-onset approximation (black lines).