

In the Era of Therapeutic Hypothermia, How Well Do Studies of Perinatal Neuroprotection Control Temperature?

Robert Galinsky^{a, b} Justin M. Dean^a Christopher A. Lear^a Joanne O. Davidson^a
Simerdeep Dhillon^a Guido Wassink^a Laura Bennet^a Alistair J. Gunn^a

^aDepartment of Physiology, Faculty of Medical and Health Sciences, University of Auckland, Auckland, New Zealand; ^bThe Ritchie Centre, Hudson Institute of Medical Research, Clayton, VIC, Australia

Keywords

Perinatal brain injury · Cerebral palsy · Hypoxic-ischemic encephalopathy · Asphyxia · Neuroprotection

Abstract

In the era of therapeutic hypothermia, reliable preclinical studies are integral to successfully identify neuroprotective strategies to further improve outcomes of encephalopathy at term. We reviewed preclinical neuroprotection studies reported between January 2014 and June 2016 to assess the use of effective temperature monitoring and control. As a secondary measure, we examined whether studies addressed other methodological issues such as stage of brain development, sex differences, the timing of the treatment relative to the insult, and the histological and functional endpoints used after hypoxia-ischemia. The extent and duration of temperature monitoring was highly inconsistent. Only a minority of papers monitored core (19/61; 31%) or brain temperature (3/61; 5%). Most (40/45) of the neuroprotectants either were likely to affect thermoregulation or their impact is unknown. In 85% of papers neonatal rodents were used (67% at P7); 51% of papers did not report the sex of the animals or tested the effect of potential neuroprotectants on just one sex. In 76% of studies, treatment was before or im-

mediately after the insult (within the first 2 h), and few studies assessed long-term histological and behavioral outcomes. In conclusion, many recent preclinical neonatal studies cannot exclude the possibility that apparent neuroprotection might be related to drug-induced hypothermia or to other methodological choices. Close monitoring and control of brain temperature during, as well as for many days after, experimental hypoxia-ischemia are now critical to reliably develop new ways to improve neurodevelopmental outcomes after perinatal hypoxic-ischemic encephalopathy.

© 2016 S. Karger AG, Basel

Introduction

Mild “therapeutic” hypothermia for hypoxic-ischemic encephalopathy (HIE) in near-term and term infants is now well established to improve survival without disability [1]. Current hypothermia protocols are only partially protective, such that nearly half of infants still die or survive with disability despite cooling [2], and so it is vital to identify adjunct therapies to further improve outcomes from HIE [3]. The corollary of the successful clinical translation of therapeutic hypothermia is that we can now be certain that even small changes in brain and/or body

temperature critically affect outcome in both animals [4–7] and humans [8–10]. Indeed, many apparent neuroprotective effects of various pharmacological agents can be mediated by drug-induced hypothermia, as previously reviewed [11]. Conversely, neuroprotection can be masked by delayed hyperthermia [6].

Neonatal rodents are widely used to model HIE and assess neuroprotective strategies. However, due to their small body mass relative to surface area, lack of subcutaneous fat, naked skin, poor control of peripheral vasculature, and absent shivering [12], the infant rat produces less heat and loses more body heat than adults [13]. These factors make them functionally poikilothermic and particularly susceptible to rapid changes in body and brain temperature during changes in environmental temperature [14]. Further, their brains are relatively small and flat, which can allow selective brain cooling, particularly if blood flow is reduced after HI [11]. Behavioral thermoregulation in neonatal rodents begins to emerge during the 1st week of life, primarily through metabolism of brown adipose tissue and group huddling [15]. However, their thermoregulatory ability is not fully developed until the 3rd week of life [16].

Thus, in this study, we reviewed studies reported between January 2014 and June 2016 to assess the use of effective temperature monitoring and control in preclinical neuroprotection studies in the era of therapeutic hypothermia. As a secondary measure, we examined whether these studies addressed other important methodological issues [17], such as the stage of brain development, sex differences, the timing of the treatment relative to the insult, and the duration of follow-up for histological and functional endpoints after HI.

Analysis Strategy

In this review, a group of researchers focusing on perinatal neuroprotection have examined preclinical studies of perinatal neuroprotection reported from January 1, 2014, to June 30, 2016, for how well these studies control for temperature in near-term or term-equivalent brain injury. The studies were grouped into those that demonstrated improved or unaffected neuropathological or neurodevelopmental outcomes. Studies were further subdivided by species, type of insult used to induce HIE, the treatment used, timing of treatment relative to the insult, extent of temperature monitoring, sex of the subjects, main study outcomes (histological and/or functional), and survival time after the insult based on histological

and/or functional assessments. Studies were searched for on PubMed, MEDLINE (OvidSP), and Google Scholar using the following terms: (brain injury OR hypoxia-ischemia OR hypoxia OR ischemia OR cerebral ischemia OR occlusion) AND (neuroprotection) AND (fetal OR neonatal OR postnatal). Other sources used to identify studies included relevant papers and reviews.

Abstracts were initially identified and screened by an investigator from our study group (R.G.) and duplicated by another investigator (J.M.D.). Full-text articles were then obtained using Papers software (version 3.4.6; Labtiva Inc., Cambridge, MA, USA) for review. Relevant studies were selected based on group consensus. Studies were deemed eligible if they presented clear histological and/or functional outcomes from in vivo experiments that investigated neuroprotection using pharmacological interventions or mechanisms of protection via less pragmatic interventions, such as gene deletion, gene inactivation, or conditioning. All pharmacological interventions were considered individually, even in studies that used more than 1 treatment. The class of drug was searched for to assess whether the neuroprotectant had potential to affect thermoregulation. Specifically, the neuroprotectants were searched for on PubMed, MEDLINE (OvidSP), and Google Scholar using the following terms: (*name of relevant drug*) AND (temperature OR vasodilation). The drug was considered *likely* to affect temperature if it had previously been shown to affect thermoregulation or belonged to a family of drugs known to affect body or brain temperature. It was deemed *unlikely* to affect temperature if no effect had previously been reported. *Unknown* indicates no studies have investigated whether the drug affects temperature. Histological outcomes were based on assessment of gray and/or white matter cell survival or death. Functional outcomes were based on assessment of behavior or recovery of brain activity, as assessed using neurophysiology or examination of clinical seizures.

Inclusion criteria were in vivo studies of rodents on postnatal days (P) 7–11, sheep at 0.80–0.85 of gestation and P1–5 neonatal piglets. These developmental ages are often used to study treatments for term infants, although the cortical maturity of laboratory rats at P7 is consistent with the late preterm human infant [18]. We excluded in vitro studies and those that did not meet the age criteria. The methodological quality of the studies was assessed according to whether temperature was controlled, treatment was randomized, and investigators were blinded to the intervention during histological and functional assessments. The results are summarized in Table 1 and Figures 1–4.

Table 1. Preclinical studies of perinatal neuroprotection

Reference	Species	Insult	Treatment (route)	Timing	Temperature, °C	Pathology	Functional outcome	Survival	Sex
<i>Improved neurodevelopmental outcomes</i>									
39	P7 mice	HI: 90 min 8% O ₂	<i>Hydrogen (i.p.)</i> HI + vehicle HI + hydrogen-rich saline	Post-Tx: 24, 48, 72 h	Ambient: 36°C, during 2 h recovery	↓ infarct vol, <i>p</i> < 0.05 vs. vehicle		3 days	?
40	P1 piglets	HI FiO ₂ 6%	HT and argon HI + HT HI + HT + argon	Post-Tx: 2 h	Core: 33.5°C, continuous monitoring	~67% ↓ cell death, <i>p</i> < 0.05 vs. HT	↑ PCr/Pi and ↓ thalamic, white matter Lac/NAA and improved aEEG recovery vs. HT, <i>p</i> < 0.05	2 days	♂
41	P1–2 piglets	HI FiO ₂ 10%	<i>Cannabidiol (i.p.) and HT</i> HI + HT + vehicle HI + HT + cannabidiol	Post-Tx: 30 min	Core: 33–34°C, continuous monitoring	↓ cortical neuronal necrosis vs. HT, <i>p</i> < 0.05	↑ Lac/NAA and improved aEEG recovery vs. HT, <i>p</i> < 0.05	6 h	♂
42	P7 rats	HI: 90 min 8% O ₂	<i>Sevoflurane</i> HI HI + sevoflurane	Immediate Tx: 0 h	Ambient: 36.5°C, during HI	↓ CA1 and 3 hippo- campal neuronal loss, <i>p</i> < 0.05 vs. HI	Improved learning and memory, <i>p</i> < 0.05 vs. HI	3 weeks	♂/♀
43	P7 rats	HI: 90 min 8% O ₂	<i>Helium</i> HI He + HI	Pre-Tx: –24 h	Ambient: 37°C, during HI	↓ infarct vol vs. HI, <i>p</i> < 0.05	Improved sensorimotor function vs. HI	Pathology: 1 day behavior: 3 weeks	?
44	P10 rats	HI: 3.5 h 8% O ₂	<i>MicroRNA-210</i> <i>antagonist (i.c.v.)</i> vehicle + HI miRNA antagonist + HI	Post-Tx: 4 h	Ambient: 37°C, during HI	47% ↓ infarct vol vs. vehicle, <i>p</i> < 0.05	Improved spatial learning and sensorimotor function vs. vehicle	Pathology: 2 days behavior: 6 weeks	?
45	P10 mice	HI: 15 min 8% O ₂	<i>DHA (i.p.)</i> HI + vehicle HI + DHA	Immediate Tx: 0, 1 h	Ambient: 37°C, during HI	49% ↓ in infarct vol, <i>p</i> < 0.05 vs. vehicle		1 day	?
45	P10 mice	HI: 15 min 8% O ₂	HI + vehicle HI + DHA	Immediate Tx: 0, 1 h	Ambient: 37°C, during HI	32% ↑ in brain tissue vol, <i>p</i> < 0.05 vs. vehicle	Improved memory performance, <i>p</i> < 0.05 vs. vehicle	9 weeks	?
46	P7 rats	HI: 2 h 8% O ₂	HT and <i>NAC (i.p.)</i> HI + HT + vehicle HI + HT + NAC	Immediate Tx: 0 h	Core: 36.5–37.5 vs. 33.5–34.5°C during 2 h recovery	↓ infarct vol in females, <i>p</i> < 0.05 HT + NAC vs. HT no effect in males	Improved sensorimotor function vs. HT.	Pathology: 2 days behavior: 6 weeks	♂/♀
47	P9 mice	HI: 1 h 10% O ₂	HT HI + NT HI + HT	Immediate Tx: 0 h	Core: 35 vs. 27°C during 4 h recovery	↑ hippocampal neuronal density and ↓ total neural injury score	Improved sensorimotor function and cognition vs. NT	Pathology: 1 week behavior: 3 weeks	♂/♀
47	P9 mice	HI: 1 h 10% O ₂	HI + NT HI + HT	Post-Tx: 2 h	Core: 35 vs. 27°C during 4 h recovery	↑ hippocampal neuronal density and ↓ neural injury score	No effect on behavior	Pathology: 1 week behavior: 5 weeks	♂/♀
48	P7 rats	HI: 2 h 8% O ₂	<i>Dexmedetomidine</i> (i.p.) HI + vehicle HI + dexmedetomidine	Post-Tx: 30, 60, and 120 min	Ambient: 37°C, during HI		Improved sensorimotor function and learning and memory vs. vehicle	4 weeks	?
49	P9 mice	HI: 50 min 10% O ₂	<i>Atg7 (autophagy- related protein 7)</i> WT + HI <i>Atg7</i> KO + HI	Constitutive gene KO	Ambient: 36°C, during HI	~49% ↓ in infarct vol, <i>p</i> < 0.05 vs. WT		8 days	♂/♀
50	P7 rats	HI: 2 h 8% O ₂	<i>Isoflurane</i> HI HI + isoflurane	Immediate Tx: 0 h	Ambient: 37°C, during HI	↑ cortical and thalamic neuronal density vs. HI	Improved learning and memory performance vs. HI	Pathology: 1 week behavior: 5 weeks	♂/♀
50	P7 rats	HI: 2 h 8% O ₂	HI HI + isoflurane	Post-Tx: 3 or 6 h	Ambient: 37°C, during HI	↑ cortical and thalamic neuronal density vs. HI	Improved learning and memory performance vs. HI	Pathology: 1 week behavior: 5 weeks	♂/♀
51	P7 rats	HI: 90 min 8% O ₂	<i>Argon</i> HI + nitrogen with O ₂ HI + argon with O ₂	Immediate Tx: 0 h	Not specified	47% ↓ in infarct vol, <i>p</i> < 0.05 vs. nitrogen		4 weeks	?
52	P7 rats	HI: 2 h 8% O ₂	<i>Taurine (i.p.)</i> HI + vehicle HI + taurine	Post-Tx: 12, 24, 26, 48 h	Ambient: 37°C, during HI	↓ infarct vol and cell death score		2 days	♂/♀
53	P10 mice	HI: 25 min 8% O ₂	<i>Creatine</i> (dietary suppl.) HI + normal diet HI + 3% creatine	Post-Tx: 10–70 days	Core: 36°C, during HI	↓ infarct vol in creatine: 17.5 ± 2.5% vs. normal diet: 27.8 ± 4.1%, <i>p</i> < 0.05	Improved memory and motor function	10 weeks	♀
54	P1 piglets	HI FiO ₂ 12%	HT HI + NT (38.5°C) HI + HT (35°C) HI + HT (33.5°C) HI + HT (30°C)	Post-Tx: 2 h	Core: 38.5 vs. 35, 33.5, or 30°C, continuous monitoring	↓ white matter apoptosis in 35 and 33.5°C groups vs. NT, <i>p</i> < 0.05	None	2 days	♂

Table 1 (continued)

Reference	Species	Insult	Treatment (route)	Timing	Temperature, °C	Pathology	Functional outcome	Survival	Sex
55	P7 rats	HI: 2 h 15 min 8% O ₂	<i>Resveratrol</i> (<i>i.p.</i>) vehicle + HI resveratrol + HI	Pre-Tx: -10 min	Ambient: 36°C, during HI	~83% ↓ infarct vol, ↓ cortical and hippocampal injury scores and ↑ myelination vs. vehicle, $p < 0.05$	Improved memory performance	Pathology: 1 week behavior: 12 weeks	♂/♀
31	P10 rats	HI: 45 min 8% O ₂	HT HI + NT HI + 4 h HT	Immediate Tx: 0 h	Core: 31 vs. 36°C during 4 h recovery	Greater ↓ in regional injury in males vs. females $p < 0.05$	None	1 week (8 days)	♂/♀
31	P10 rats	HI: 45 min 8% O ₂	HI + NT HI + 4 h HT	Immediate Tx: 0 h	Core: 31 vs. 36°C during 4 h recovery	↓ cortical and hippocampal injury in males; no significant effect of HT in females	HT improved working memory in males but not females	Pathology and behavior 3 weeks (20 days)	♂/♀
56	P10 rats	HI: 2.5 h 8% O ₂	<i>G-CSF</i> (<i>s.c.</i>) HI + vehicle HI + <i>G-CSF</i>	Post-Tx 1 h	Ambient: 37°C, during HI	34% ↓ infarct vol vs. vehicle, $p < 0.05$	None	1 day	?
57	P7 mice	HI: 100 min 7.5% O ₂	<i>Carvacrol</i> (<i>i.p.</i>) vehicle + HI carvacrol + HI (30 or 50 mg/kg)	Pre-Tx: -30 min	Ambient: 37°C, during HI	↓ infarct vol by 46 and 90% in 30- and 50-mg groups, respectively vs. vehicle, $p < 0.05$	Improved sensorimotor performance in 50 mg/kg vs vehicle, $p < 0.05$	Pathology: 1 day behavior: 1 week	?
58	P10 mice	HI: 25 min 8% O ₂	<i>Creatine monohydrate</i> (<i>dietary suppl.</i>) HI + normal diet HI + 2% creatine	Post-Tx: 10–105 days	Ambient: 36°C, during HI	No change in infarct vol vs. normal diet, $p < 0.05$	Improved motor function, learning and memory vs. normal diet	Pathology: 15 weeks behavior: 17 weeks (118 days)	♂
59	P7 rats	HI: 2.5 h 8% O ₂	<i>Uridine</i> (<i>i.p.</i>) HI + vehicle HI + uridine	Immediate Tx: 0 h, 1, and 2 days	Ambient: 37°C, during HI	↓ infarct vol: uridine, 11 ± 1% vs. vehicle, 20 ± 3%, $p < 0.05$	None	3 days	♂/♀
60	P10 mice	HI: 45 min 8% O ₂	HT HI + NT HI + 4 h HT	Immediate Tx: 0 h	Ambient: 37 vs. 28.5°C during 4 h of recovery	~34% ↑ NeuN+ staining area vs. HI + NT, $p < 0.05$	None	1 week	♂
61	P7 rats	HI: 2.5 h 8% O ₂	<i>Gap26</i> (<i>Cx43 mimetic peptide</i>) (<i>i.p.</i>) vehicle + HI Gap26 + HI	Pre-Tx: -1 h	Ambient: 37°C, during HI	↓ infarct vol by ~60% vs. vehicle, $p < 0.05$		2 days	?
61	P7 rats	HI: 2.5 h 8% O ₂	<i>Gap26</i> (<i>Cx43 mimetic peptide</i>) (<i>i.p.</i>) HI + vehicle HI + Gap26	Post-Tx: 24 h then daily for 7 days	Ambient: 37°C, during HI	None	Improved motor function and memory vs. vehicle	3 weeks	?
62	P7 rats	HI: 90 min 8% O ₂	HT and Xe HI + NT HI + 5 h HT HI + 5 h HT + 50% Xe	Post-Tx: 5 h	Core: 36°C, during HI 37 vs. 32°C, during 5 h of recovery	No additive effect of Xe on infarct area vs. HT, $p > 0.05$	HT + Xe improved sensorimotor performance vs. HT; negative correlation between brain injury and motor function females ($r = -0.76$, $p < 0.05$) not males ($p = 0.06$)	Pathology: 10 weeks behavior: 9 weeks	♂/♀
63	P7 rats	HI: 150 min 8% O ₂	<i>Vitexin</i> (5, 7, 4-trihydroxy-flavone-8-glucoside) (<i>i.p.</i>) HI + vehicle HI + vitexin	Post-Tx: 5 min	Ambient: 37°C, during HI	~64% ↓ infarction volume and ↓ cortical and hippocampal neuronal cell death, $p < 0.05$ vs. vehicle	None	2 days	♂/♀
29	P10–11 rats	HI: 1 h 8% O ₂	HT HI + NT HI + 4 h HT	Immediate Tx: 0 h	Core: 36–36.5 vs. 32°C, during 4 h of recovery	At week 12: ↓ total and regional damage scores vs. HI + NT, $p < 0.05$	HT improved motor function in moderate but not severely injured subjects vs. NT, $p < 0.05$	Pathology: 12 weeks behavior: 9 weeks	♂/♀
64	P7 rats	HI: 120 min 8% O ₂	<i>NAC</i> (<i>i.p.</i>) vehicle + HI NAC + HI	Pre-Tx: -30 min, then daily from P8 to P44	Ambient: 36°C, during HI	↑ MBP expression in corpus callosum vs. vehicle, $p < 0.05$	Improved sensorimotor function vs. vehicle	4 weeks	♂
65	P7 rats	HI 3.5 h 8% O ₂	<i>Progesterone</i> (<i>i.p. and s.c.</i>) HI + vehicle HI + progesterone	Immediate Tx: 0 h <i>i.p.</i> then 2 h and 1, 2, 3, 4, 5, 6, and 7 days <i>s.c.</i>	Ambient: 37°C, during HI	30% ↓ mean tissue loss in HI + progesterone males vs. vehicle (no effect in females)	Behavioral tests showed no benefit of progesterone vs. vehicle, $p > 0.05$	Pathology: 7 weeks behavior: 6 weeks	♂/♀
66	P7 rats	HI: 2 h 8% O ₂	HT NT during HI HT during HI	During HI	Ambient: 36.1 vs. 32.2°C during 2 h HI, 35.7–37.0°C during recovery	None	Greater improvement in sensory motor and memory performance in HT females vs. HT males vs. NT	10–11 weeks (73–76 days)	♂/♀

Table 1 (continued)

Reference	Species	Insult	Treatment (route)	Timing	Temperature, °C	Pathology	Functional outcome	Survival	Sex
67	P10 mice	HI: 15 min 8% O ₂	<i>Isoflurane</i> HI + room air HI + isoflurane	Immediate Tx: 0 h	Ambient: 37–37.5°C, during HI	37% ↓ infarct vol vs. room air	Improved memory performance vs. room air	Pathology: ? 1 day behavior: 8 weeks	?
68	P7–9 mice	HI: 60 min 7.5% O ₂	<i>HPC and K_{ATP} channel blocker (tolbutamide)</i> (i.p.) HI HPC + tolbutamide + HI HPC + vehicle + HI	Pre-Tx: –2 days	Core: 37°C, during HI and 30 min of recovery	↓ infarct vol in HPC + vehicle: 21.4 ± 3.5% vs. HPC + tolbutamide: 45.3 ± 1.7%, <i>p</i> < 0.05; similar infarct vol between HI and HPC + tolbutamide	None	1 day	♂/♀
68	P7–9 mice	HI: 100 min 7.5% O ₂	HI HPC + vehicle + HI HPC + tolbutamide + HI	Pre-Tx: 2 days	Core: 37°C, during HI and 30 min of recovery	↓ infarct vol in HPC + vehicle by ~45 and 40% vs. HI and HPC + tolbutamide, respectively, <i>p</i> < 0.05	Improved sensorimotor performance in HPC vs. HPC + tolbutamide, <i>p</i> < 0.05	Pathology and behavior: 1 week	♂/♀
69	P7 rats	HI: 2.5 h 8% O ₂	<i>HT</i> HI + NT HI + 3 h HT	Immediate Tx: 0 h	Core: 38 vs. 30°C, during 3 h of recovery	None	Improved neurobehavioral scores vs. NT	3 h	♂
70	P7 rats	HI: 2.5 h 8% O ₂	<i>P5-TAT</i> (i.p.) vehicle + HI P5-TAT + HI (1, 10, 25, 50 µg/kg)	Pre-Tx: –1 h	Ambient: 37°C, during HI	Dose-dependent ↓ infarct vol. 58% ↓ in 50 µg/kg group, <i>p</i> < 0.05	None	2 days	?
70	P7 rats	HI: 2.5 h 8% O ₂	HI + vehicle HI + P5-TAT (50 µg/kg/day)	Post-Tx: 1–7 days after HI	Ambient: 37°C, during HI	None	Improved sensorimotor performance vs. vehicle	4 weeks	?
71	P8 mice	HI: 30 min 10% O ₂	<i>NPI</i> (neuronal pentraxin 1) WT + HI NPI KO + HI	Constitutive gene knockout	Ambient: 35°C, during 2 h recovery	↑ cortical, striatal, and hippocampal vol by ~45, 50 and 50%, respectively, <i>p</i> < 0.05 vs. WT	None	1 week	?
72	P7 rats	HI: 2 h 8% O ₂	<i>iNOS inhibition</i> (i.c.v.) vehicle + HI iNOS inhibitor + HI	Pre-Tx: –4 days	Ambient: 37°C, during HI	↑ hippocampal neuronal density by ~67 and 64% in CA1 and CA3, respectively. <i>p</i> < 0.05 vs. vehicle	Improved motor function, learning and memory, <i>p</i> < 0.05 vs. vehicle	Pathology: 1 week behavior: 3 weeks	♂/♀
73	P7 rats	HI: 150 min 8% O ₂	<i>OMT</i> (oxymatrine) (i.p.) vehicle + HI OMT + HI	Pre-Tx: –15 min then every 12 h for 2 days post-HI	Ambient: 37°C, during HI	~55% ↓ infarct area and ↓ neuronal cell death vs. vehicle, <i>p</i> < 0.05	Improved sensorimotor function and memory vs vehicle, <i>p</i> < 0.05	Pathology: 2 days behavior: 7 weeks	♂
74	P9 mice	LPS + HI 20 min 10% O ₂	<i>IDR-1018</i> (innate defense regulator peptide 1018) (i.p.) LPS + HI + vehicle LPS + HI + IDR-1018	Post-Tx: 3 h	Not specified	↓ brain injury score in cortex, thalamus hippocampus and striatum and ↓ white matter loss by ~80%, <i>p</i> < 0.05 vs. vehicle	None	1 week	♂/♀
75	P7 rats	HI: 2 h 8% O ₂	<i>Sildenafil</i> (i.p.) HI + vehicle HI + sildenafil citrate	Immediate Tx: 0 h	Not specified	↓ brain tissue loss by 43 ± 15% and ↑ myelination in cingulum and external capsule, <i>p</i> < 0.05 vs vehicle	Intermediate improvement in motor performance vs. vehicle	1 week	♂/♀
76	P10 rats	HI: 120 min 8% O ₂	<i>Dexamethasone</i> (i.c.v.) vehicle + HI dexamethasone + HI	Pre-Tx: immediately before HI	Not specified	~57% ↓ infarction size vs. vehicle	None	2 days	♂/♀
77	P9 mice	HI: 50 min 10% O ₂	<i>Q-VD-OPh</i> (broad-spectrum caspase inhibitor) (i.p.) HI + vehicle HI + Q-VD-OPh	Post-Tx: 12 h then every 24 h for 2 weeks	Ambient: 36°C, during HI	↓ tissue loss (by 31.3%), decreased neuropathologi- cal scores in cortex, hippocampus, and thalamus vs. vehicle, <i>p</i> < 0.05	Improved sensorimotor function vs. vehicle, <i>p</i> < 0.05	Behavior: 6 weeks pathology: 16 weeks	♂
78	P7 rats	HI: 2 h 8% O ₂	<i>nNOS</i> (7-nitroindazole) or <i>iNOS</i> (aminoguanidine) inhibition (i.p.) vehicle + HI 7-nitroindazole + HI aminoguanidine + HI	Pre-Tx: 30 min	Not specified	Complete and partial reduction in cortical infarct vol in nNOS and iNOS inhibitor groups, respectively, vs. vehicle, <i>p</i> < 0.05	None	1 week	♂
78	P7 rats	HI: 2 h 8% O ₂	HI + vehicle HI + 7-nitroindazole HI + aminoguanidine	Post-Tx: 3 h	Not specified	Partial reduction in cortical infarct vol in iNOS group vs. nNOS and vehicle, <i>p</i> < 0.05	None	1 week	♂
79	Fetal sheep (0.85 GA)	5 × 10 min UCO	<i>Allopurinol</i> (maternal) (i.v.) UCO + vehicle UCO + allopurinol	Tx during UCO	Not specified	↓ hippocampal (CA3 and CA4) neuronal damage score vs. vehicle	None	2 days	?

Table 1 (continued)

Reference	Species	Insult	Treatment (route)	Timing	Temperature, °C	Pathology	Functional outcome	Survival	Sex
80	P7 rats	HI: 120 min 8% O ₂	<i>Caffeine citrate</i> (i.p.) caffeine + HI vehicle + HI	Pre-Tx: immediately before HI then 0, 24, 48, and 72 h	Ambient: 37°C, during HI	↓ cell death in the hippocampus and parietal cortex vs. vehicle, $p < 0.05$	None	1 week	?
81	P7 rats	HI: 2.5 h 8% O ₂	<i>29D7 mAb</i> (<i>tropomyosin related kinase B agonist</i>) (i.c.v.) vehicle + HI 29D7 (0.1 nmol) + HI 29D7 (0.3 nmol) + HI	Pre-Tx: immediately before HI	Core: 35–37°C, measured intermittently for 7 days post-HI	↓ % area tissue loss in striatum, hippocampus and cortex $p < 0.05$ 0.1 and 0.3 nmol vs. vehicle, $p < 0.05$	None	1 week	♂/♀
81	P7 rats	HI: 2.5 h 8% O ₂	vehicle + HI 29D7 (0.3 nmol) + HI	Pre-Tx: immediately before HI	Core: 35–37°C, measured intermittently for 7 days post-HI	↓ % area tissue loss in striatum, hippocampus and cortex by ~72, 77 and 77%, respectively, $p < 0.05$ vs. vehicle	Improved sensory motor function vs. vehicle, $p < 0.05$	5 weeks	♂/♀
82	P7 mice	HI: 30 min 8% O ₂	<i>Cl-amidine</i> (<i>peptidyl- arginine deiminase inhibition</i>) (i.p.) LPS + HI + vehicle LPS + HI + Cl-amidine	Immediate Tx: 0 h	Ambient: 36°C, during HI	↓ infarct vol in the hippocampus and external capsule by ~62 and 74%, respectively, $p < 0.05$	None	2 days	?
83	P10 mice	HI: 60 min 10% O ₂	<i>HT and sevoflurane</i> HI + 10% O ₂ 1 h HI + 3.5% sevoflurane + HT (1 h)	Immediate Tx: 0 h	Core: ~34 vs. 36.5–37.5°C during HI	↓ neuronal damage score in neocortex, striatum, thalamus, and hippocampus vs. HI + 10% O ₂ , $p < 0.05$	Improved learning and memory vs. HI + 10% O ₂ , $p < 0.05$	Pathology: 1 week behavior: 8 weeks	♂/♀
84	P7 rats	HI: 90 min 8% O ₂	<i>HT and Xe</i> HI + NT HI + 5 h HT HI + 5 h HT + Xe 50%	Immediate Tx: 0 h	Core: 36°C during HI, 37 vs. 32°C, during 5 h of recovery	↓ infarct area in HI + HT + Xe (19%) vs. HI + HT (37%) and NT (54%), $p < 0.05$	None	1 week	♂/♀
85	P7 rats	HI: 2 h 8% O ₂	<i>Ia1P</i> (<i>inter-α inhibitor protein</i>) (i.p.) vehicle + HI human Ia1P + HI	Pre-Tx: immediately before and 24 h after HI	Not specified	Intermediate ↑ cortical and hippocampal tissue vol. vs. vehicle	Improved spatial and nonspatial learning vs. vehicle	Pathology: 22 weeks (157 days) behavior: 11 weeks (80 days)	♂
86	P7 rats	HI: 55 min 8% O ₂	<i>Estrol</i> (<i>estrogenic steroid</i>) (i.p.) vehicle + HI estrol 1, 5, 10, 50 mg/kg/ day + HI	Pre-Tx: –3 days	Ambient: 37°C, during HI	↓ infarct area in all treatment groups vs. vehicle, $p < 0.05$	None	1 week	?
86	P7 rats	HI: 55 min 8% O ₂	HI + vehicle HI + estrol 1, 5, 10, 50 mg/kg	Immediate Tx: 0 h	Ambient: 37°C, during HI	↓ infarct area in all treatment groups vs. vehicle	None	1 week	?
87	P8 rats	HI: 45 min 7.7% O ₂	<i>Lithium</i> (i.p.) HI + vehicle HI + lithium	Post-Tx: 5–19 days	Ambient: 36°C, during HI	↓ tissue vol loss 169 ± 26 vs. 277 ± 27 mm ³ and pathological score, $p < 0.05$ vs. vehicle	Improved motor- and ↓ anxiety- related activity, $p < 0.05$ vs. vehicle	Pathology: 12 weeks behavior: 6 weeks	♂
88	P7 rats	HI: 2 h 8% O ₂	<i>Isoflurane or cyclosporine A</i> (<i>mitochondrial permeability transition pore inhibitor</i>) (i.c.v.) HI + vehicle HI + isoflurane HI + cyclosporine A	Immediate Tx: 0 h	Ambient: 37°C, during HI	↑ hippocampal and thalamic neuronal density in isoflurane and cyclosporine A groups vs. vehicle, $p < 0.05$		1 week	♂/♀
88	P7 rats	HI: 2 h 8% O ₂	HI + vehicle HI + isoflurane HI + cyclosporine A	Immediate Tx: 0 h	Ambient: 37°C, during HI	↑ hippocampal and thalamic neuronal density in isoflurane and cyclosporine A groups vs. vehicle, $p < 0.05$	Similar improvement in spatial learning and memory in isoflurane and cyclosporine A groups vs. vehicle	5 weeks	♂/♀
89	P3–5 piglets	45 min FiO ₂ 10% then asphyxia 7 min	<i>HT and HETE-0016</i> (<i>20-hydroxyeicosatetraenoic acid inhibitor</i>) (i.v.) Hypox + NT Hypox + HT Hypox + HETE-0016 + HT	Post-Tx: HETE-0016 at 5 min HT at 3 h	Core: 38.5–39.5 vs. 34.0°C, continuous monitoring	↑ neuronal survival in cortex (~22%), putamen (~14%), and thalamus (~28%) with combination therapy, $p < 0.05$ vs. HT	No effect on time to feed after extubation; no significant effect on n of clinical seizures, $p < 0.1$	1 week (10 days)	♂

Table 1 (continued)

Reference	Species	Insult	Treatment (route)	Timing	Temperature, °C	Pathology	Functional outcome	Survival	Sex
<i>No improvement in neurodevelopmental outcomes</i>									
90	P7 rats	HI: 150 min 8% O ₂	HT and Xe NT HT HT + Xe	Immediate Tx: 0 h	Core: 37 vs. 32°C, during 5 h of recovery	No effect of HT or HT + Xe on infarct vol and neuronal loss vs. NT		1 week	♂/♀
55	P7 rats	HI: 2 h 15 min 8% O ₂	Resveratrol (<i>i.p.</i>) HI + vehicle HI + resveratrol	Immediate Tx: 0 h	Ambient: 36°C, during HI	No effect on infarct vol, neuronal loss or myelination vs. vehicle	None	1 week	♂/♀
91	P9 mice	HI: 50 min 10% O ₂	Osteopontin (<i>i.n./i.p.</i>) HI + vehicle HI + osteopontin	Immediate and post-Tx: 0, and 3 h and 1–3 days <i>i.n.</i> or 0 h and every 2 days on days 1–15 <i>i.p.</i> or 0 h and every 2 days on days 5–15 <i>i.p.</i>	Ambient: 37°C, during HI	No effect on white (MBP) or gray (MAP2) matter area in any treatment group vs. vehicle; no sex-specific effect	No effect on sensorimotor performance	Pathology: 9 weeks (65 days) behavior: 8 weeks (60 days)	♂/♀
23	Fetal sheep (0.85 GA)	Bilateral carotid artery occlusion 30 min	HT ischemia + NT ischemia + HT 3 days ischemia + HT 5 days	Post-Tx: 3 h	Brain: 39.5 vs. 31.3– 31.8°C, continuous monitoring	↓ neuronal survival in cortex and dentate gyrus in 5- vs. 3-h HT group, <i>p</i> < 0.05	Improved EEG recovery and ↓ brain edema in 3- and 5-day groups, <i>p</i> < 0.05 vs. NT	1 week	♂/♀
22	Fetal sheep (0.85 GA)	Bilateral carotid artery occlusion 30 min	HT and Cx43 (<i>connexin</i> <i>hemichannel blockade</i>) (<i>i.c.v.</i>) ischemia + NT ischemia + HT ischemia + HT + Cx43	Post-Tx: 3 h	Brain: 39.5 vs. 31.3– 32.1°C, continuous monitoring	No additive protection with Cx43 block + HT vs. HT	No additive effect with Cx43 block + HT on EEG recovery or brain edema vs. HT	1 week	♂/♀
92	P1 piglets	45 min FiO ₂ 6%	HT (<i>selective head cooling</i>) Hypox + NT Hypox + HT	Immediate Tx: 0 h	Brain: 38.5 vs. 31°C, continuously for 24 h then daily	No improvement in neuropathology scores in cortical and subcortical gray or white matter	No difference in <i>n</i> of electrographic or clinical seizures between groups	3 days	♂/♀
63	P7 rats	HI: 150 min 8% O ₂	Vitexin (5, 7, 4-trihydroxyflavone-8- glucoside) (<i>i.p.</i>) HI + vehicle HI + vitexin	Post-Tx: 3 h	Ambient: 37°C, during HI	No effect on infarct vol vs. vehicle	None	2 days	♂/♀
93	P7 rats	LPS + HI: 50 min 8% O ₂	HT LPS + HI + NT LPS + HI + 5 h HT	Immediate Tx: 0 h	Core: 37 vs. 32°C, during 5 h of recovery	No effect on hippocampal neuronal loss vs. NT	None	1 week	♂/♀
94	P3–5 piglets	HI 45 min 10% O ₂ then asphyxia 7 min	HT HI + NT HI + HT HI + HT slow rewarming HI + HT fast rewarming	Post-Tx: 2 h	Core: 38.5–39.5 vs. 34.0°C, continuous monitoring	↑ cortical neuronal apoptosis in both rewarming groups vs. HT and NT groups, <i>p</i> < 0.05	None	1 day	♂
95	P7 rats	LPS + HI 50 min 8% O ₂	HT LPS + HI + 5 h NT LPS + HI + 5 h HT	Immediate Tx: 0 h	Core: 36°C during HI 37 vs. 32°C during 5 h of recovery	No effect on % brain area loss, HT vs. NT	None	1 week	♂/♀
84	P7 rats	HI: 90 min 8% O ₂	HI + NT HI + 5 h HT 35°C HI + 5 h HT 35°C + Xe 20%	Post-Tx: 4 h	Core: 36°C, during HI 37 vs. 32°C, during 5 h of recovery	No effect of delayed combination or HT alone on infarct area vs. NT	None	1 week	♂/♀

aEEG, amplitude-integrated electroencephalogram; DHA, docosahexaenoic acid; GA, gestational age; G-CSF, granulocyte colony stimulating factor; HT, hypothermia; HI, hypoxia-ischemia; Hypox: hypoxia; HPC, hypoxic preconditioning; iNOS, inducible nitric oxide synthase; LPS, lipopolysaccharide; MBP, myelin basic protein; NAA, N-acetyl aspartate; NAC, N-acetyl-L-cysteine; NeuN, neuronal nuclear protein; NT, normothermia; nNOS: neuronal nitric oxide synthase; P, postnatal day; P5-TAT, cycline-dependent kinase 5 inhibitor; Tx, treatment; UCO, umbilical cord occlusion; vol, volume; WT, wild type; Xe, xenon.

Studies of Perinatal Neuroprotection

We identified and screened a total of 171 papers. One hundred and ten were excluded due to inappropriate developmental age, *ex vivo* assessment, or lack of assessment of neurological outcomes. Thus 61 individual pa-

pers were included in this analysis (Fig. 1). Studies were further subdivided by treatment regime (i.e., studies that investigated more than 1 timing for treatment relative to the insult) or where pathological and functional outcomes were assessed at different survival times. After

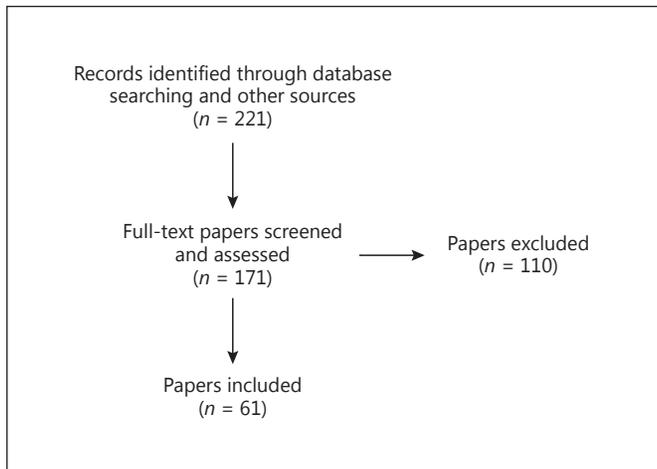


Fig. 1. Flow chart illustrating the number of papers identified through database searching and other relevant sources, the number of full-text articles screened, assessed, and excluded, and the final number of original papers surveyed.

subdividing the studies that used more than 1 survival time for pathological and functional assessments or assessed outcomes after more than 1 treatment time, we identified 75 individual studies. For the purpose of reporting on the treatment regime, timing, outcomes, and survival, we summarized the data based on the individual studies (total $n = 75$). As each of the subdivided studies used the same developmental age at the time of the insult and protocols for temperature control and documentation of sex, the developmental age, type and duration of temperature monitoring, and reporting of sex are summarized based on the number of original papers (total $n = 61$).

The majority of papers 52/61 (85%) studied neonatal rats or mice; 35 out of these 52 (67%) papers studied neonatal rodents at P7, the remainder studied rodents between P8 and P11 (17/52; 32%). These ages are broadly comparable to the late preterm or term human infant at 32–40 weeks [17–19]. All studies used the Rice-Vannucci model of unilateral carotid artery ligation followed by a period of moderate hypoxia [18]. A range of pathological assessments of injury was reported, such as measurements of total and/or regional infarct area or volume or semiquantitative assessments to assign a neuronal injury score. Functional outcomes were assessed using a wide variety of behavioral tests examining memory, sensorimotor function, coordination, or neophobia.

Nine out of 61 papers (15%) studied large animals. Histopathological assessments were mainly performed

by quantifying the total number of viable or dying cells within regions susceptible to injury. In term-equivalent fetal sheep, injury was induced using bilateral carotid artery occlusion or repeated umbilical cord occlusion. In the piglet, injury was induced using prolonged isocapnic hypoxia with or without a short (7-min) period of severe asphyxia or bilateral carotid artery occlusion. In both paradigms, injury has been reported in “watershed” zones (areas of white and gray matter that lie within the borders between major arteries where perfusion pressure is least), such as the parasagittal cortex, the dorsal horn of the hippocampus, thalamus, dentate gyrus, cerebellar neocortex, striatum, and parasagittal and periventricular white matter. Functional outcomes were assessed using electroencephalography (EEG) to evaluate recovery of activity and/or electrographic or clinical seizure assessment. In neonatal piglets, clinical seizures were also assessed. In fetal sheep, cortical impedance was used to examine the degree of cytotoxic edema after the insult.

Results

Temperature Monitoring

In 7 of 61 papers, temperature monitoring was not reported in the study design or outcomes. Of these, 6 were in rodents and 1 was in fetal sheep. All of these papers reported neuroprotection (Fig. 1a). Environmental temperature only was reported in 32 papers (all in rodents); 31 out of these 32 papers reported neuroprotection, including 2 papers that tested hypothermia, and 1 paper reported no improvement in histological or functional outcomes. Twenty-eight of these 32 papers reported environmental temperature during HI only (Fig. 2a, 3a).

Core temperature was monitored in 19/61 papers (31%; 14 papers used rodents and 5 papers used piglets; Fig. 2). In rodents, 1/14 papers monitored temperature intermittently for 7 days (Fig. 3), 2 papers monitored core temperature during HI, and 2 papers monitored core temperature during HI for 30 min or 2 h afterwards; all of these papers reported improved outcomes with treatment. Nine papers in rodents monitored core temperature for 3–10 h after HI (Fig. 3c). Of these, 6/9 papers reported neuroprotection at least in some treatment groups.

In all papers using piglets, core temperature was monitored continuously (Fig. 3b). All of these papers focused on hypothermia strategies or adjuvant therapy with hy-

Fig. 2. Number of papers that did not monitor temperature (none) and papers that reported ambient (environmental), core, and brain temperature (T) in rodents (**a**) and large animals (**b**) from 61 papers investigating neuroprotection for term/near-term HIE between January 1, 2014, and June 30, 2016. Open bars show the number of studies that reported no protection. Black bars show the number of studies that reported neuroprotection.

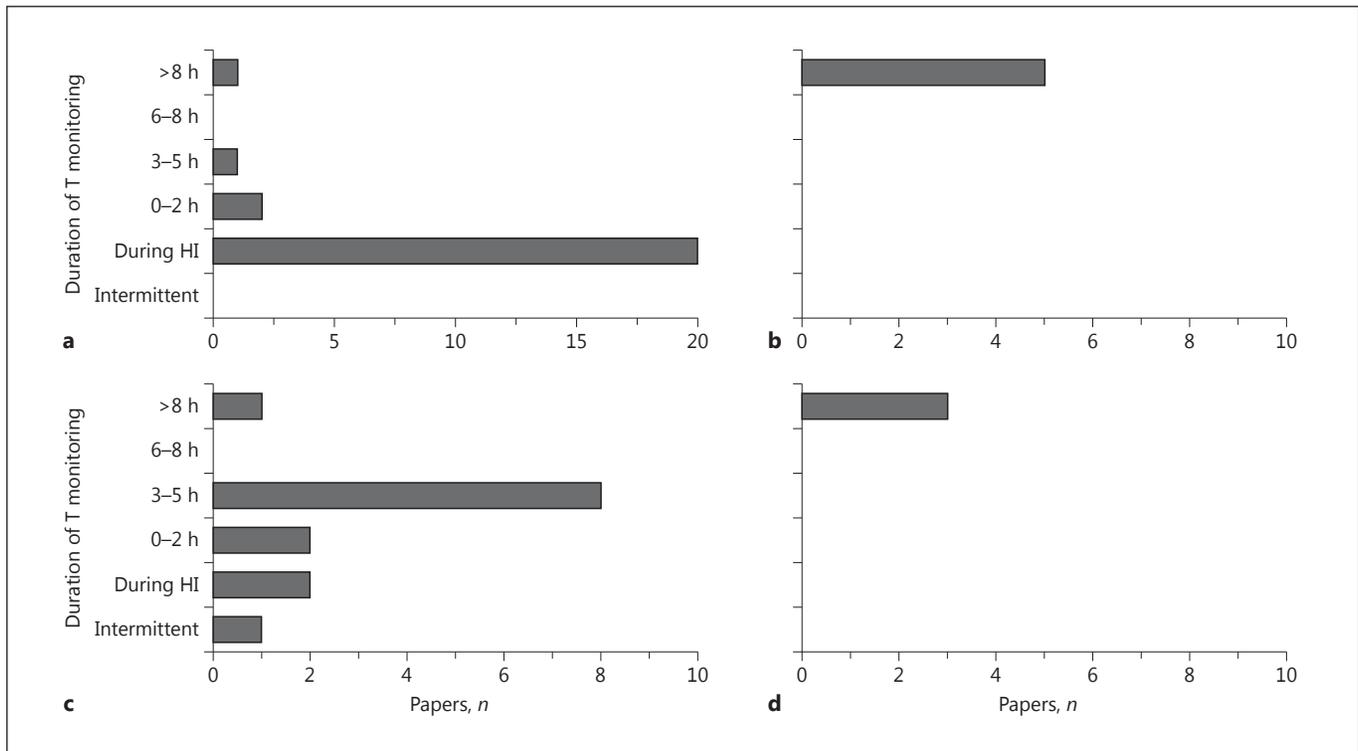
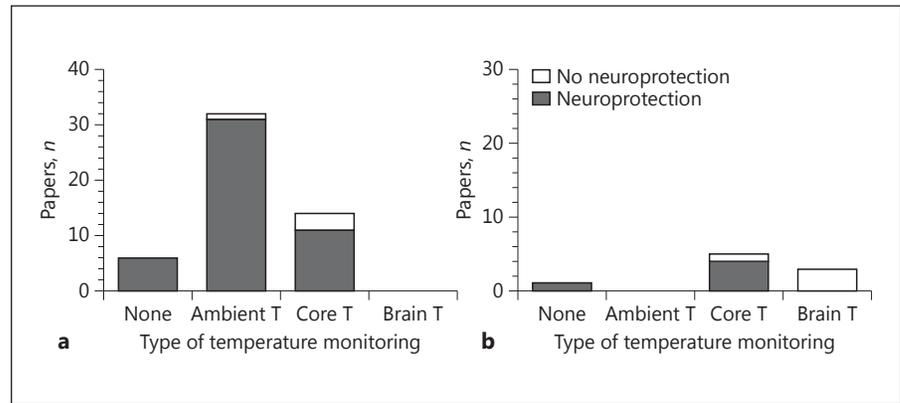


Fig. 3. Number of papers in rodents ($n = 39$; **a**, **c**) and large animals ($n = 7$; **b**, **d**) that reported the duration of temperature (T) monitoring: ambient/environmental (**a**) and core temperature (**c**) monitoring in rodents, and core (**b**) and brain temperature (**d**) monitoring in large animals. Total $n = 46$.

pothermia, and all reported at least partial protection. Brain temperature was monitored in 3/61 papers (5%; 1 in piglets and 2 in fetal sheep; Fig. 3d). In the piglet study, brain temperature was monitored for 24 h and then once daily for 2 days. In 2 papers using fetal sheep, extradural brain temperature was monitored continuously throughout the experiment.

Possibility that Neuroprotectants Affected Thermoregulation

We identified a total of 45 neuroprotectants/protective strategies from the papers reviewed in this study. Twenty treatments were classified as likely to affect thermoregulation, and 5 were considered unlikely to affect temperature control. The effect on thermoregulation was unknown in 20 cases (Fig. 4a).

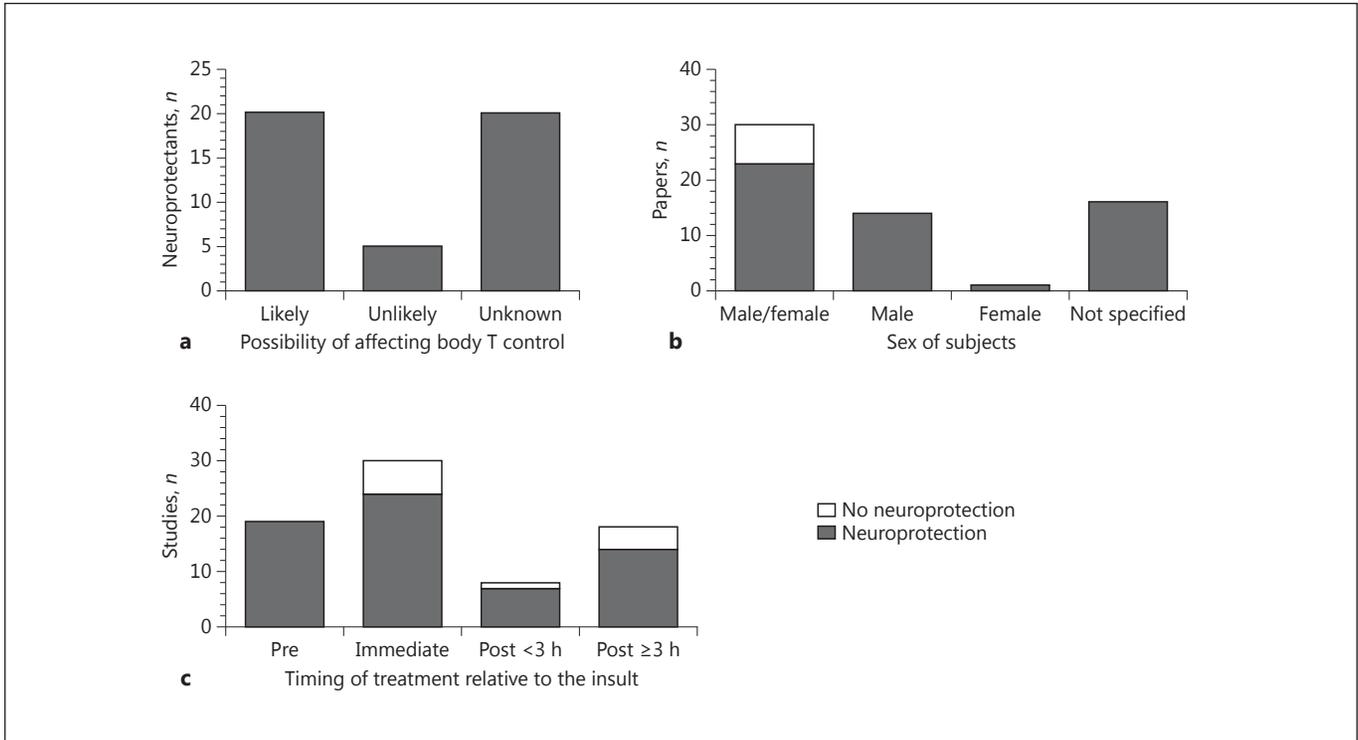
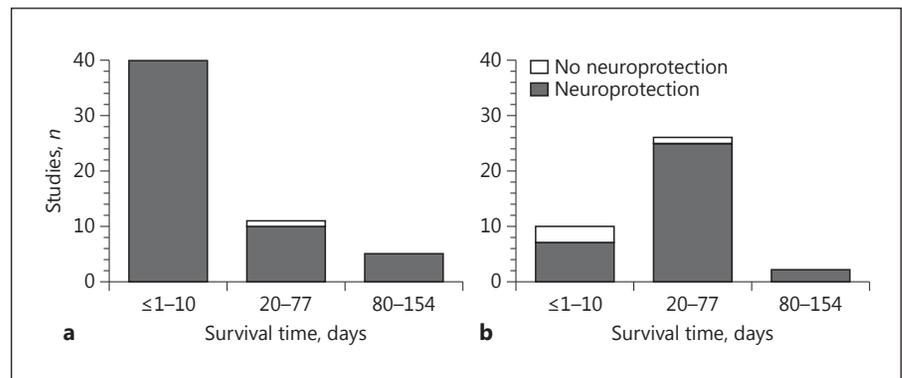


Fig. 4. a Probability of neuroprotectants affecting thermoregulation; 36 neuroprotectants were identified and assessed. *Likely* was defined as a drug previously shown to affect temperature (T) or belonging to a family of drugs reported to affect temperature. *Unlikely* refers to a drug previously shown to have no effect on temperature. *Unknown* indicates no studies have investigated whether the drug/neuroprotective strategy affects temperature. **b** Number

of papers that reported no neuroprotection (white bars) or neuroprotection (black bars) in either both sexes, males, or females, or did not report the sex of the subjects studied. **c** Number of studies that reported no neuroprotection (white bars) or neuroprotection (black bars) after treating before, immediately, within less than 3 h, or greater than or equal to 3 h after the insult.

Fig. 5. Number of studies that reported no protection (white bars) or neuroprotection (black bars) after using short ($\leq 1-10$ days), medium (20–77 days), or long (80–154 days) survival times for histological (**a**) and behavioral assessment (**b**).



Route of Drug Delivery

The majority of the papers surveyed (52/61; 85%) assessed the effectiveness of therapeutic candidates without hypothermia. Nine out of 61 papers (15%) investigated hypothermia with adjuvant therapies. Twelve out of 61 papers (20%) investigated hypothermia alone. The treat-

ment regimens for candidate drugs were highly variable. Pharmacological interventions were administered in either single or multiple doses. Most of the rodent studies administered candidate treatments subcutaneously or intra-peritoneally ($n = 32$). One rodent study used a combination of intraperitoneal and intra-nasal drug delivery,

and 12 studies delivered candidate drugs in gaseous form during or after HI. Eight studies (7 in rodents and 1 in fetal sheep) used intracerebroventricular drug delivery, 2 studies (both in rodents) delivered the drug as a dietary supplement, 1 study in fetal sheep administered the candidate drug via the maternal circulation, and 1 study in neonatal piglets used intravenous infusion. Two rodent studies used a constitutive gene knockout.

Sex

Thirty of 61 papers (49%) examined the effect of treatment on both sexes (Fig. 4b). Twenty-three studies showed improved neural outcomes, and 7 studies showed no improvement. In a further 14 papers, therapeutic candidates were examined in males, and 1 paper reported outcomes in females only. All reported improved neural outcomes. Finally, in 16 papers, the sex of the subjects was not reported; all of these studies reported neuroprotection.

Timing of Treatment and Neurodevelopmental Outcomes

In total, 64/75 studies demonstrated neuroprotection for HIE. In 19 studies, treatment began before the start of HI (Fig. 4c), including two studies of constitutive gene knockouts (i.e. before HI). In 31 studies, treatment was delivered during or immediately after HI or within 2 h. In just 14 studies, treatment was started 3 h or later after HI. The survival time before histological assessment in 44 studies reporting neuroprotection was less than or equal to 1–10 days (Fig. 5a). In 10 studies, histological outcomes were reported between 20 and 77 days. In 5 studies, outcomes were reported between 80 and 154 days. Five studies did not report histological outcomes.

Thirty-four studies (32 in rodents and 2 in piglets) reported improved functional outcomes. Of these, 7 assessed functional outcomes after less than or equal to 1–10 days, 25 after 20–77 days, and 2 after 80–154 days (Fig. 5b). Interestingly, in 5 studies, histological and functional outcomes were discordant, such that 3 studies reported improved histology with no improvement in function, and 2 studies found improved functional outcomes without improved histology.

Eleven out of 75 perinatal studies reported no neuroprotection with treatment. Seven studies began treatment immediately after HI (or within the first 2 h), and 4 studies delayed treatment for up to 3–4 h after HI (Fig. 4c). In 10 of the 11 studies, histological analysis was performed after relatively short survival times of less than or equal to 1–10 days. One study in rodents examined histological

outcomes after approximately 9 weeks (65 days; Fig. 5a). Four studies assessed functional outcomes. Three studies (1 in piglets and 2 in fetal sheep) assessed functional outcomes based on EEG or clinical seizures after less than or equal to 1–10 days, and 1 study in rodents examined behavioral outcomes after 60 days (Fig. 5b).

Discussion

This systematic review shows that the majority of recent studies investigating perinatal neuroprotection for HIE between 2014 and 2016 were performed in rodents, of which only a minority rigorously monitored and controlled body and brain temperatures. Thus, many recent studies cannot exclude the possibility that their findings could have been confounded by unappreciated changes in brain temperature due to drug-induced cooling or environmental conditions.

Carefully conducted studies in neonatal rodents offer many advantages for studies of neuroprotection [17]. They are easy to use and cost effective, which in turn facilitates data collection from large cohorts at multiple time points. Further, rodent studies enable wide application of histological techniques, molecular biology probes, and behavioral assessments, allowing investigators to provide detailed mechanistic and functional insight. However, as highlighted in this systematic review, considerable care is required to be certain that iatrogenic changes in brain temperature do not occur. Large animal studies are expensive, require more laboratory infrastructure relative to studies in smaller animals, and offer limited opportunities for assessing neurodevelopmental outcomes other than neurophysiological monitoring. Nevertheless, they are easier to monitor and have intrinsically greater thermal stability.

The great majority of recent papers used neonatal mice or rats (53/61). Only 14 papers studying rodents measured body temperature at all, and just 9 continuously monitored body temperature for short periods of 3–10 h after HI. No studies reported on temperatures after this early phase of recovery. Given the near-poikilothermic nature of P7 neonatal rats and mice, environmental temperature is often used as a surrogate for body temperatures. However, although most of the studies measured environmental temperature during the insult, few monitored or reported temperature after HI. This is an important consideration since even normal, healthy neonatal rodents often show lower body temperatures in the typical laboratory nesting conditions between P7

and P14 [20]. Core temperature in neonatal rodents will be affected by the amount and composition of nesting material, position in the nest, huddling, distance from the dam, and the time since the last period of suckling. Conversely, the majority of large animal studies continuously monitored and controlled brain and or core temperature throughout the study period. One study did not report temperature in near-term fetal sheep; however, a major advantage of testing potential neuroprotectants in fetal sheep is that body temperature is regulated by the pregnant ewe and, therefore, unless the ewe is febrile, fetal temperatures are highly stable [21–24].

In adult rodents, the use of anesthetics such as halothane, sodium pentobarbital, and isoflurane around the time of the insult have all been shown to induce hypothermia, as previously reviewed [11]. Furthermore, many rodent studies have reported that the beneficial effects of potential neuroprotectants were confounded by hypothermia during the first 6–8 h after the insult [11, 25] and that maintenance of normothermia abolishes neuroprotection [25]. In this study, the majority (40/45; 89%) of the pharmacological interventions being tested over the last 2 years belonged to classes of drugs that are either likely to affect body temperature or whose effect on thermoregulation is unknown. Collectively, these data strongly suggest that significant age-related and drug-induced fluctuations in body temperature, which might affect the severity of neural injury and, therefore, could confound studies of perinatal neuroprotection, may have been missed in the majority of recent studies.

The relative maturity of the species being studied is important for understanding the potential clinical application of the studies. The brain maturation of near-term fetal sheep (0.8–0.85 of gestation) and neonatal piglets is considered to be consistent with term human infants [26, 27]. However, at P7, the rat is closest to the 32- to 36-week-old human infant based on the immaturity of the germinal zone, the limited extent of cortical layering, and reduced density of synapses in the striate cortex [18, 28]. The majority of recent reports of neuroprotection in rodents (64%) studied P7 rats. As recently reviewed, neonatal rats are closest to term equivalent at P10–P14 [17]. The responses to HI at P10 have been carefully characterized [29]. Even at P10–P14, healthy neonatal rodents often show significant spontaneous central hypothermia in the typical laboratory nesting environment [20], and therefore they still require continuous core temperature monitoring and control.

About half (31/61; 51%) of the papers surveyed in this review reported outcomes either for only one sex or did not specify the sex of the subjects used. Preclinical studies have reported sexual dimorphisms in the severity and evolution of injury after HI and in responses to treatment, as previously reviewed [30]. For example, in P7 rats, males exposed to HI had greater deficits in some but not all behavioral tasks compared to females [30]. Similarly, there is some evidence of more variable responses to cooling in female mice [31]. It remains unclear whether there are similar differences in humans [32]. However, these data demonstrate the importance of carefully testing therapeutic agents in both sexes, as recommended by the National Institutes of Health [33].

A key clinical-translation consideration for testing potential neuroprotectants is when to treat. The majority of studies surveyed (57/75; 76%) started intervention either before or immediately after the insult. Scientifically, since the mechanisms of intra-insult treatment are different from those after the insult cell death, efficacy during pretreatment does not necessarily suggest a benefit after the insult. Clinically, because most HI occurs around birth, it is very difficult to identify fetuses who are likely to develop postnatal HIE in advance as the positive predictive value of heart rate monitoring in labor is exceedingly low [34]. It remains a formidable challenge to start any intervention even within the first 6 h after birth [35]. Thus, in most cases, it is very difficult to translate preclinical neuroprotective agents until the window of opportunity *after* HI is known. Collectively, these data support evaluation of delayed treatment after the insult as part of routine preclinical evaluation.

It is still important to test the effectiveness of potential neuroprotectants during normothermia to help understand the mechanism/s by which they induce neuroprotection and to evaluate their potential for use in patients who may not be suitable candidates for therapeutic hypothermia. Nevertheless, given that hypothermia is now standard clinical practice [35, 36], ultimately it is essential to also test the effectiveness of potential drug therapies as adjuvants to therapeutic hypothermia in order to improve current treatment protocols. In this analysis, just 9 out of the 61 papers surveyed (15%) assessed the effectiveness of hypothermia with adjuvant therapies.

Finally, the majority of studies (54/75; 72%) examined relatively short survival times, from ≤ 1 to 10 days. Less than half of the studies reported functional outcomes after treatment. Shorter survival times provide crucial information about the early outcome of injury and any intervention. Equally, particularly after moderate insults,

injury can continue to evolve over many weeks [37], and behavioral and histological outcomes are sometimes discordant, as noted in this analysis.

This paper examined a sample of preclinical publications investigating neuroprotection between 2014 and 2016. It is not possible to know whether the methodological issues identified in our analysis affected the outcome of the studies. Nevertheless, the findings reported in this study are highly consistent with previous reports of the “adult” literature [38]. If this critical information is not presented in the publications, it is not possible to assess the significance of past and future studies in a meaningful way. Finally, we did not examine papers assessing mechanisms of HIE. It is very likely that these variables are just as important to understand the mechanisms of HIE and, therefore, it would be well worth analyzing them in the future.

Conclusion

The majority of recent studies of perinatal neuroprotection reported limited, or even no, assessment or maintenance of brain or body temperature. In addition, many did not address pragmatic treatment regimes, report or test the sex of the subjects, or assess long-term functional and histological outcomes. It is critical that all future preclinical studies of the mechanisms and treatment of perinatal HIE should carefully control brain and core temperatures during and after HI.

References

- 1 Jacobs SE, Berg M, Hunt R, Tarnow-Mordi WO, Inder TE, Davis PG: Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database Syst Rev* 2013; 1:CD003311.
- 2 Edwards AD, Brocklehurst P, Gunn AJ, Halliday H, Juszczak E, Levene M, Strohm B, Thoresen M, Whitelaw A, Azzopardi D: Neurological outcomes at 18 months of age after moderate hypothermia for perinatal hypoxic ischaemic encephalopathy: synthesis and meta-analysis of trial data. *BMJ* 2010;340: c363.
- 3 Robertson NJ, Tan S, Groenendaal F, van Bel F, Juul SE, Bennet L, Derrick M, Back SA, Valdez RC, Northington F, Gunn AJ, Mallard C: Which neuroprotective agents are ready for bench to bedside translation in the newborn infant? *J Pediatr* 2012;160:544.e4–552.e4.
- 4 Wass CT, Lanier WL, Hofer RE, Scheithauer BW, Andrews AG: Temperature changes of $> 1^\circ\text{C}$ alter functional neurologic outcome and histopathology in a canine model of complete cerebral ischemia. *Anesthesiology* 1995; 83:325–335.
- 5 Coimbra C, Boris-Moller F, Drake M, Wieloch T: Diminished neuronal damage in the rat brain by late treatment with the antipyretic drug dipyrone or cooling following cerebral ischemia. *Acta Neuropathol* 1996;92: 447–453.
- 6 Coimbra C, Drake M, Boris-Moller F, Wieloch T: Long-lasting neuroprotective effect of postischemic hypothermia and treatment with an anti-inflammatory/antipyretic drug. Evidence for chronic encephalopathic processes following ischemia. *Stroke* 1996;27: 1578–1585.
- 7 Campbell K, Meloni BP, Knuckey NW: Combined magnesium and mild hypothermia (35°C) treatment reduces infarct volumes after permanent middle cerebral artery occlusion in the rat at 2 and 4, but not 6 h. *Brain Res* 2008;1230:258–264.
- 8 Azzimondi G, Bassein L, Nonino F, Fiorani L, Vignatelli L, Re G, D’Alessandro R: Fever in acute stroke worsens prognosis. A prospective study. *Stroke* 1995;26:2040–2043.
- 9 Terent A, Andersson B: The prognosis for patients with cerebrovascular stroke and transient ischemic attacks. *Ups J Med Sci* 1981;86: 63–74.
- 10 Hindfelt B: The prognostic significance of subfebrility and fever in ischaemic cerebral infarction. *Acta Neurol Scand* 1976;53:72–79.
- 11 DeBow SB, Clark DL, MacLellan CL, Colbourne F: Incomplete assessment of experimental cytoprotectants in rodent ischemia studies. *Can J Neurol Sci* 2003;30:368–374.
- 12 Hull GC: Thermoregulation in young mammals; in Whittow GC (ed): *Comparative Physiology of Thermoregulation*. New York, Academic Press, 1973, pp 167–200.
- 13 Taylor PM: Oxygen consumption in newborn rats. *J Physiol* 1960;154:153–168.

Contributor’s Statement

Robert Galinsky, Justin M. Dean, and Alistair J. Gunn conceptualized and designed the study. Robert Galinsky performed the initial search and analysis, and drafted the paper. Christopher A. Lear, Joanne O. Davidson, Simerdeep Dhillon, Guido Wassink, Laura Bennet, and Alistair J. Gunn provided in-depth review of the studies identified in the search and revised the paper. All authors critically reviewed the paper and approved the final version as submitted and agree to be accountable for all aspects of the work.

Funding Sources

This study was supported by the Health Research Council of New Zealand, Auckland Medical Research Foundation, and National Health and Medical Research Council (RG: 1090890), and the Victorian Government’s Operational Infrastructure Support Program.

Disclosure Statement

The authors have no financial relationships relevant to this article and no conflicts of interest to disclose. The study sponsors had no role in (1) the study design; (2) the collection, analysis, and interpretation of data; (3) the writing of the report; and (4) the decision to submit the paper for publication. No honorarium, grant, or other form of payment was given to anyone to produce the paper.

- 14 Hahn P, Koldovsky O: Prenatal and postnatal development of metabolism and its regulation in mammals (in Czech). *Cesk Fysiol* 1966;215:437–479.
- 15 Farrell WJ, Alberts JR: Rat behavioral thermoregulation integrates with nonshivering thermogenesis during postnatal development. *Behav Neurosci* 2007;121:1333–1341.
- 16 Fitzgerald LR, Bacon RL: Metabolic rate and thermal regulation in neonatal mice. *Anat Rec* 1952;113:532.
- 17 Patel SD, Pierce L, Ciardiello AJ, Vannucci SJ: Neonatal encephalopathy: pre-clinical studies in neuroprotection. *Biochem Soc Trans* 2014;42:564–568.
- 18 Rice JE 3rd, Vannucci RC, Brierley JB: The influence of immaturity on hypoxic-ischemic brain damage in the rat. *Ann Neurol* 1981;9:131–141.
- 19 Romijn HJ, Hofman MA, Gramsbergen A: At what age is the developing cerebral cortex of the rat comparable to that of the full-term newborn human baby? *Early Hum Dev* 1991;26:61–67.
- 20 Wood T, Osredkar D, Puchades M, Maes E, Falck M, Flatebo T, Walloe L, Sabir H, Thoresen M: Treatment temperature and insult severity influence the neuroprotective effects of therapeutic hypothermia. *Sci Rep* 2016;6:23430.
- 21 Galinsky R, Draghi V, Wassink G, Davidson JO, Drury PP, Lear CA, Gunn AJ, Bennet L: Magnesium sulfate reduces EEG activity but is not neuroprotective after asphyxia in pre-term fetal sheep. *J Cereb Blood Flow Metab* 2016, Epub ahead of print.
- 22 Davidson JO, Rout AL, Wassink G, Yuill CA, Zhang FG, Green CR, Bennet L, Gunn AJ: Non-additive effects of delayed connexin hemichannel blockade and hypothermia after cerebral ischemia in near-term fetal sheep. *J Cereb Blood Flow Metab* 2015;35:2052–2061.
- 23 Davidson JO, Wassink G, Yuill CA, Zhang FG, Bennet L, Gunn AJ: How long is too long for cerebral cooling after ischemia in fetal sheep? *J Cereb Blood Flow Metab* 2015;35:751–758.
- 24 Laburn HP, Mitchell D, Goelst K: Fetal and maternal body temperatures measured by radiotelemetry in near-term sheep during thermal stress. *J Appl Physiol* 1992;72:894–900.
- 25 Galinsky R, Bennet L, Groenendaal F, Lear CA, Tan S, van Bel F, Juul SE, Robertson NJ, Mallard C, Gunn AJ: Magnesium is not consistently neuroprotective for perinatal hypoxia-ischemia in term-equivalent models in pre-clinical studies: a systematic review. *Dev Neurosci* 2014;36:73–82.
- 26 Dobbing J, Sands J: Comparative aspects of the brain growth spurt. *Early Hum Dev* 1979;3:79–83.
- 27 Barlow RM: The foetal sheep: morphogenesis of the nervous system and histochemical aspects of myelination. *J Comp Neurol* 1969;135:249–262.
- 28 Semple BD, Blomgren K, Gimlin K, Ferriero DM, Noble-Haesslein LJ: Brain development in rodents and humans: identifying benchmarks of maturation and vulnerability to injury across species. *Prog Neurobiol* 2013;106–107:1–16.
- 29 Patel SD, Pierce L, Ciardiello A, Hutton A, Paskewitz S, Aronowitz E, Voss HU, Moore H, Vannucci SJ: Therapeutic hypothermia and hypoxia-ischemia in the term-equivalent neonatal rat: characterization of a translational pre-clinical model. *Pediatr Res* 2015;78:264–271.
- 30 Smith AL, Alexander M, Rosenkrantz TS, Sadek ML, Fitch RH: Sex differences in behavioral outcome following neonatal hypoxia ischemia: insights from a clinical meta-analysis and a rodent model of induced hypoxic ischemic brain injury. *Exp Neurol* 2014;254:54–67.
- 31 Burnsed JC, Chavez-Valdez R, Hossain MS, Kesavan K, Martin LJ, Zhang J, Northington FJ: Hypoxia-ischemia and therapeutic hypothermia in the neonatal mouse brain – a longitudinal study. *PLoS One* 2015;10:e0118889.
- 32 Wyatt JS, Gluckman PD, Liu PY, Azzopardi D, Ballard RA, Edwards AD, Ferriero DM, Polin RA, Robertson CM, Thoresen M, Whitelaw A, Gunn AJ; CoolCap Study Group: Determinants of outcomes after head cooling for neonatal encephalopathy. *Pediatrics* 2007;119:912–921.
- 33 Wright DW, Espinoza TR, Merck LH, Ratcliff JJ, Backster A, Stein DG: Gender differences in neurological emergencies. Part II. A consensus summary and research agenda on traumatic brain injury. *Acad Emerg Med* 2014;21:1414–1420.
- 34 Nelson KB, Dambrosia JM, Ting TY, Grether JK: Uncertain value of electronic fetal monitoring in predicting cerebral palsy. *N Engl J Med* 1996;334:613–618.
- 35 Gunn AJ, Bennet L: Timing still key to treating hypoxic ischaemic brain injury. *Lancet Neurol* 2016;15:126–127.
- 36 Jacobs SE, Berg M, Hunt R, Tarnow-Mordi WO, Inder TE, Davis PG: Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database Syst Rev* 2013;1:CD003311.
- 37 Geddes R, Vannucci RC, Vannucci SJ: Delayed cerebral atrophy following moderate hypoxia-ischemia in the immature rat. *Dev Neurosci* 2001;23:180–185.
- 38 Klahr AC, Nadeau CA, Colbourne F: Temperature control in rodent neuroprotection studies: methods and challenges. *Ther Hypothermia Temp Manag* 2016, Epub ahead of print.
- 39 Bai X, Liu S, Yuan L, Xie Y, Li T, Wang L, Wang X, Zhang T, Qin S, Song G, Ge L, Wang Z: Hydrogen-rich saline mediates neuroprotection through the regulation of endoplasmic reticulum stress and autophagy under hypoxia-ischemia neonatal brain injury in mice. *Brain Res* 2016;1646:410–417.
- 40 Broad KD, Fierens I, Fleiss B, Rocha-Ferreira E, Ezzati M, Hassell J, Alonso-Alconada D, Bainbridge A, Kawano G, Ma D, Tachtsidis I, Gressens P, Golay X, Sanders RD, Robertson NJ: Inhaled 45–50% argon augments hypothermic brain protection in a piglet model of perinatal asphyxia. *Neurobiol Dis* 2016;87:29–38.
- 41 Lafuente H, Pazos MR, Alvarez A, Mohammed N, Santos M, Arizti M, Alvarez FJ, Martinez-Orgado JA: Effects of cannabidiol and hypothermia on short-term brain damage in new-born piglets after acute hypoxia-ischemia. *Front Neurosci* 2016;10:323.
- 42 Lai Z, Zhang L, Su J, Cai D, Xu Q: Sevoflurane postconditioning improves long-term learning and memory of neonatal hypoxia-ischemia brain damage rats via the PI3K/Akt-mPTP pathway. *Brain Res* 2016;1630:25–37.
- 43 Li Y, Liu K, Kang ZM, Sun XJ, Liu WW, Mao YF: Helium preconditioning protects against neonatal hypoxia-ischemia via nitric oxide mediated up-regulation of antioxidants in a rat model. *Behav Brain Res* 2016;300:31–37.
- 44 Ma Q, Dasgupta C, Li Y, Bajwa NM, Xiong F, Harding B, Hartman R, Zhang L: Inhibition of microRNA-210 provides neuroprotection in hypoxic-ischemic brain injury in neonatal rats. *Neurobiol Dis* 2016;89:202–212.
- 45 Mayurasakorn K, Niatsetskaya ZV, Sosunov SA, Williams JJ, Zirpoli H, Vlasakov I, Deckerbaum RJ, Ten VS: DHA but not EPA emulsions preserve neurological and mitochondrial function after brain hypoxia-ischemia in neonatal mice. *PLoS One* 2016;11:e0160870.
- 46 Nie X, Lowe DW, Rollins LG, Bentzley J, Fraser JL, Martin R, Singh I, Jenkins D: Sex-specific effects of N-acetylcysteine in neonatal rats treated with hypothermia after severe hypoxia-ischemia. *Neurosci Res* 2016;108:24–33.
- 47 Reinboth BS, Koster C, Abberger H, Prager S, Bendix I, Felderhoff-Muser U, Herz J: Endogenous hypothermic response to hypoxia reduces brain injury: implications for modeling hypoxic-ischemic encephalopathy and therapeutic hypothermia in neonatal mice. *Exp Neurol* 2016;283:264–275.
- 48 Ren X, Ma H, Zuo Z: Dexmedetomidine post-conditioning reduces brain injury after brain hypoxia-ischemia in neonatal rats. *J Neuroimmune Pharmacol* 2016;11:238–247.
- 49 Xie C, Ginet V, Sun Y, Koike M, Zhou K, Li T, Li H, Li Q, Wang X, Uchiyama Y, Truttmann AC, Kroemer G, Puyal J, Blomgren K, Zhu C: Neuroprotection by selective neuronal deletion of Atg7 in neonatal brain injury. *Autophagy* 2016;12:410–423.
- 50 Xu Y, Xue H, Zhao P, Yang Y, Ji G, Yu W, Han G, Ding M, Wang F: Isoflurane postconditioning induces concentration- and timing-dependent neuroprotection partly mediated by the GluR2 AMPA receptor in neonatal rats after brain hypoxia-ischemia. *J Anesth* 2016;30:427–436.

- 51 Zhao H, Mitchell S, Ciechanowicz S, Savage S, Wang T, Ji X, Ma D: Argon protects against hypoxic-ischemic brain injury in neonatal rats through activation of nuclear factor (erythroid-derived 2)-like 2. *Oncotarget* 2016;7:25640–25651.
- 52 Zhu XY, Ma PS, Wu W, Zhou R, Hao YJ, Niu Y, Sun T, Li YX, Yu JQ: Neuroprotective actions of taurine on hypoxic-ischemic brain damage in neonatal rats. *Brain Res Bull* 2016;124:295–305.
- 53 Allah Yar R, Akbar A, Iqbal F: Creatine monohydrate supplementation for 10 weeks mediates neuroprotection and improves learning/memory following neonatal hypoxia ischemia encephalopathy in female albino mice. *Brain Res* 2015;1595:92–100.
- 54 Alonso-Alconada D, Broad KD, Bainbridge A, Chandrasekaran M, Faulkner SD, Kerenyi A, Hassell J, Rocha-Ferreira E, Hristova M, Fleiss B, Bennett K, Kelen D, Cady E, Gressens P, Golay X, Robertson NJ: Brain cell death is reduced with cooling by 3.5°C to 5°C but increased with cooling by 8.5°C in a piglet asphyxia model. *Stroke* 2015;46:275–278.
- 55 Arteaga O, Revuelta M, Uriguén L, Alvarez A, Montalvo H, Hilario E: Pretreatment with resveratrol prevents neuronal injury and cognitive deficits induced by perinatal hypoxia-ischemia in rats. *PLoS One* 2015;10:e0142424.
- 56 Charles MS, Drunalini Perera PN, Doycheva DM, Tang J: Granulocyte-colony stimulating factor activates JAK2/PI3K/PDE3B pathway to inhibit corticosterone synthesis in a neonatal hypoxic-ischemic brain injury rat model. *Exp Neurol* 2015;272:152–159.
- 57 Chen W, Xu B, Xiao A, Liu L, Fang X, Liu R, Turlova E, Barszczyk A, Zhong X, Sun CL, Britto LR, Feng ZP, Sun HS: TRPM7 inhibitor carvacrol protects brain from neonatal hypoxic-ischemic injury. *Mol Brain* 2015;8:11.
- 58 Iqbal S, Ali M, Iqbal F: Long term creatine monohydrate supplementation, following neonatal hypoxic ischemic insult, improves neuromuscular coordination and spatial learning in male albino mouse. *Brain Res* 2015;1603:76–83.
- 59 Koyuncuoglu T, Turkyilmaz M, Goren B, Cetinkaya M, Cansev M, Alkan T: Uridine protects against hypoxic-ischemic brain injury by reducing histone deacetylase activity in neonatal rats. *Restor Neurol Neurosci* 2015;33:777–784.
- 60 Kwak M, Lim S, Kang E, Furmanski O, Song H, Ryu YK, Mintz CD: Effects of neonatal hypoxic-ischemic injury and hypothermic neuroprotection on neural progenitor cells in the mouse hippocampus. *Dev Neurosci* 2015;37:428–439.
- 61 Li X, Zhao H, Tan X, Kostrzewa RM, Du G, Chen Y, Zhu J, Miao Z, Yu H, Kong J, Xu X: Inhibition of connexin43 improves functional recovery after ischemic brain injury in neonatal rats. *Glia* 2015;63:1553–1567.
- 62 Liu X, Dingley J, Scull-Brown E, Thoresen M: Adding 5 h delayed xenon to delayed hypothermia treatment improves long-term function in neonatal rats surviving to adulthood. *Pediatr Res* 2015;77:779–783.
- 63 Min JW, Hu JJ, He M, Sanchez RM, Huang WX, Liu YQ, Bsoul NB, Han S, Yin J, Liu WH, He XH, Peng BW: Vitexin reduces hypoxia-ischemia neonatal brain injury by the inhibition of HIF-1 α in a rat pup model. *Neuropharmacology* 2015;99:38–50.
- 64 Park D, Shin K, Choi EK, Choi Y, Jang JY, Kim J, Jeong HS, Lee W, Lee YB, Kim SU, Joo SS, Kim YB: Protective effects of N-acetyl-L-cysteine in human oligodendrocyte progenitor cells and restoration of motor function in neonatal rats with hypoxic-ischemic encephalopathy. Evidence-based complementary and alternative medicine: eCAM 2015;2015:764251.
- 65 Peterson BL, Won S, Geddes RI, Sayeed I, Stein DG: Sex-related differences in effects of progesterone following neonatal hypoxic brain injury. *Behav Brain Res* 2015;286:152–165.
- 66 Smith AL, Garbus H, Rosenkrantz TS, Fitch RH: Sex differences in behavioral outcomes following temperature modulation during induced neonatal hypoxic ischemic injury in rats. *Brain Sci* 2015;5:220–240.
- 67 Sosunov SA, Ameer X, Niatsetskaia ZV, Utkina-Sosunova I, Ratner VI, Ten VS: Isoflurane anesthesia initiated at the onset of reperfusion attenuates oxidative and hypoxic-ischemic brain injury. *PLoS One* 2015;10:e0120456.
- 68 Sun HS, Xu B, Chen W, Xiao A, Turlova E, Alibraham A, Barszczyk A, Bae CY, Quan Y, Liu B, Pei L, Sun CL, Deurloo M, Feng ZP: Neuronal K(ATP) channels mediate hypoxic preconditioning and reduce subsequent neonatal hypoxic-ischemic brain injury. *Exp Neurol* 2015;263:161–171.
- 69 Takenouchi T, Sugiura Y, Morikawa T, Nakanishi T, Nagahata Y, Sugioka T, Honda K, Kubo A, Hishiki T, Matsuura T, Hoshino T, Takahashi T, Suematsu M, Kajimura M: Therapeutic hypothermia achieves neuroprotection via a decrease in acetylcholine with a concurrent increase in carnitine in the neonatal hypoxia-ischemia. *J Cereb Blood Flow Metab* 2015;35:794–805.
- 70 Tan X, Chen Y, Li J, Li X, Miao Z, Xin N, Zhu J, Ge W, Feng Y, Xu X: The inhibition of Cdk5 activity after hypoxia/ischemia injury reduces infarct size and promotes functional recovery in neonatal rats. *Neuroscience* 2015;290:552–560.
- 71 Thatipamula S, Al Rahim M, Zhang J, Hosain MA: Genetic deletion of neuronal pentraxin 1 expression prevents brain injury in a neonatal mouse model of cerebral hypoxia-ischemia. *Neurobiol Dis* 2015;75:15–30.
- 72 Wang Z, Feng C, Zhao H, Ren X, Peng S, Zuo Z: Autoregulation of inducible nitric oxide synthase expression by RNA interference provides neuroprotection in neonatal rats. *Theranostics* 2015;5:504–514.
- 73 Zhao P, Zhou R, Li HN, Yao WX, Qiao HQ, Wang SJ, Niu Y, Sun T, Li YX, Yu JQ: Oxy-matine attenuated hypoxic-ischemic brain damage in neonatal rats via improving antioxidant enzyme activities and inhibiting cell death. *Neurochem Int* 2015;89:17–27.
- 74 Bolouri H, Savman K, Wang W, Thomas A, Maurer N, Dullaghan E, Fjell CD, Ek CJ, Hagberg H, Hancock RE, Brown KL, Mallard C: Innate defense regulator peptide 1018 protects against perinatal brain injury. *Ann Neurol* 2014;75:395–410.
- 75 Charriaut-Marlangue C, Nguyen T, Bonnin P, Duy AP, Leger PL, Csaba Z, Pansiot J, Bourgeois T, Renolleau S, Baud O: Sildenafil mediates blood-flow redistribution and neuroprotection after neonatal hypoxia-ischemia. *Stroke* 2014;45:850–856.
- 76 Gonzalez-Rodriguez PJ, Li Y, Martinez F, Zhang L: Dexamethasone protects neonatal hypoxic-ischemic brain injury via L-PGDS-dependent PGD2-DP1-pERK signaling pathway. *PLoS One* 2014;9:e114470.
- 77 Han W, Sun Y, Wang X, Zhu C, Blomgren K: Delayed, long-term administration of the caspase inhibitor Q-VD-OPH reduced brain injury induced by neonatal hypoxia-ischemia. *Dev Neurosci* 2014;36:64–72.
- 78 Hsu YC, Chang YC, Lin YC, Sze CI, Huang CC, Ho CJ: Cerebral microvascular damage occurs early after hypoxia-ischemia via nNOS activation in the neonatal brain. *J Cereb Blood Flow Metab* 2014;34:668–676.
- 79 Kaandorp JJ, Derks JB, Oudijk MA, Torrance HL, Harmsen MG, Nikkels PG, van Bel F, Visser GH, Giussani DA: Antenatal allopurinol reduces hippocampal brain damage after acute birth asphyxia in late gestation fetal sheep. *Reprod Sci* 2014;21:251–259.
- 80 Kilicdag H, Daglioglu YK, Erdogan S, Zorludemir S: Effects of caffeine on neuronal apoptosis in neonatal hypoxic-ischemic brain injury. *J Matern Fetal Neonatal Med* 2014;27:1470–1475.
- 81 Kim GS, Cho S, Nelson JW, Zipfel GJ, Han BH: TrkB agonist antibody pretreatment enhances neuronal survival and long-term sensory motor function following hypoxic ischemic injury in neonatal rats. *PLoS One* 2014;9:e88962.
- 82 Lange S, Rocha-Ferreira E, Thei L, Mawjee P, Bennett K, Thompson PR, Subramanian V, Nicholas AP, Peebles D, Hristova M, Raivich G: Peptidylarginine deiminases: novel drug targets for prevention of neuronal damage following hypoxic ischemic insult (HI) in neonates. *J Neurochem* 2014;130:555–562.
- 83 Lin EP, Miles L, Hughes EA, McCann JC, Vorhees CV, McAuliffe JJ, Loepke AW: A combination of mild hypothermia and sevoflurane affords long-term protection in a modified neonatal mouse model of cerebral hypoxia-ischemia. *Anesth Analg* 2014;119:1158–1173.
- 84 Sabir H, Walloe L, Dingley J, Smit E, Liu X, Thoresen M: Combined treatment of xenon and hypothermia in newborn rats – additive or synergistic effect? *PLoS One* 2014;9:e109845.

- 85 Threlkeld SW, Gaudet CM, La Rue ME, Dugas E, Hill CA, Lim YP, Stonestreet BS: Effects of inter-alpha inhibitor proteins on neonatal brain injury: age, task and treatment dependent neurobehavioral outcomes. *Exp Neurol* 2014;261:424–433.
- 86 Tskitishvili E, Nisolle M, Munaut C, Pequeux C, Gerard C, Noel A, Foidart JM: Estetrol attenuates neonatal hypoxic-ischemic brain injury. *Exp Neurol* 2014;261:298–307.
- 87 Xie C, Zhou K, Wang X, Blomgren K, Zhu C: Therapeutic benefits of delayed lithium administration in the neonatal rat after cerebral hypoxia-ischemia. *PLoS One* 2014;9:e107192.
- 88 Zhao P, Ji G, Xue H, Yu W, Zhao X, Ding M, Yang Y, Zuo Z: Isoflurane postconditioning improved long-term neurological outcome possibly via inhibiting the mitochondrial permeability transition pore in neonatal rats after brain hypoxia-ischemia. *Neuroscience* 2014; 280:193–203.
- 89 Zhu J, Wang B, Lee JH, Armstrong JS, Kulikowicz E, Bhalala US, Martin LJ, Koehler RC, Yang ZJ: Additive neuroprotection of a 20-HETE inhibitor with delayed therapeutic hypothermia after hypoxia-ischemia in neonatal piglets. *Dev Neurosci* 2015;37:376–389.
- 90 Sabir H, Osredkar D, Maes E, Wood T, Thoresen M: Xenon combined with therapeutic hypothermia is not neuroprotective after severe hypoxia-ischemia in neonatal rats. *PLoS One* 2016;11:e0156759.
- 91 Bonestroo HJ, Nijboer CH, van Velthoven CT, van Bel F, Heijnen CJ: The neonatal brain is not protected by osteopontin peptide treatment after hypoxia-ischemia. *Dev Neurosci* 2015;37:142–152.
- 92 Hoque N, Liu X, Chakkarapani E, Thoresen M: Minimal systemic hypothermia combined with selective head cooling evaluated in a pig model of hypoxia-ischemia. *Pediatr Res* 2015; 77:674–680.
- 93 Osredkar D, Sabir H, Falck M, Wood T, Maes E, Flatebo T, Puchades M, Thoresen M: Hypothermia does not reverse cellular responses caused by lipopolysaccharide in neonatal hypoxic-ischaemic brain injury. *Dev Neurosci* 2015;37:390–397.
- 94 Wang B, Armstrong JS, Lee JH, Bhalala U, Kulikowicz E, Zhang H, Reyes M, Moy N, Spicer D, Zhu J, Yang ZJ, Koehler RC, Martin LJ, Lee JK: Rewarming from therapeutic hypothermia induces cortical neuron apoptosis in a swine model of neonatal hypoxic-ischemic encephalopathy. *J Cereb Blood Flow Metab* 2015;35:781–793.
- 95 Osredkar D, Thoresen M, Maes E, Flatebo T, Elstad M, Sabir H: Hypothermia is not neuroprotective after infection-sensitized neonatal hypoxic-ischemic brain injury. *Resuscitation* 2014;85:567–572.